

Ecology, diversity and seasonal distribution of wild mushrooms in a Nigerian tropical forest reserve

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Manuscript received: 19 December 2017. Revision accepted: 19 January 2018.

Abstract. Adeniyi M, Odeyemi Y, Odeyemi O. 2018. Ecology, diversity and seasonal distribution of wild mushrooms in a Nigerian tropical forest reserve. *Biodiversitas* 19: 285-295. This study investigated the ecology, diversity and seasonal distribution of wild mushrooms at Environmental Pollution Science and Technology (ENPOST) forest reserve, Ilesa, Southwestern Nigeria. Mushrooms growing in the ligneous and terrestrial habitats of the forest were collected, identified and enumerated between March 2014 and March 2015. Diversity indices including species richness, dominance, and species diversity were evaluated. Correlation ($p < 0.05$) was determined among climatic data and diversity indices. A total of 151 mushroom species specific to their respective habitats were obtained. The highest monthly species richness (70) was obtained in October 2014. While a higher dominance was observed in the terrestrial habitat during the rainy and dry seasons (0.072 and 0.159 respectively), species diversity was higher in the ligneous and terrestrial habitats during the rainy season (3.912 and 3.304 respectively). Overall, the highest carpophores in ligneous and terrestrial habitats were recorded in *Schizophyllum commune* (10,737) and *Mycena monticola* (760) correspondingly. Correlation analysis revealed that average monthly precipitation positively correlated with the relative abundance of mushrooms in the terrestrial habitat ($r = 0.716$, $p = 0.013$). This study shows the diversity of mushrooms at ENPOST forest, thereby necessitating strict and sustainable conservation measures especially those with great economic values.

Keywords: ENPOST forest, ecology, diversity indices, wild mushrooms, Nigeria

INTRODUCTION

Adequate knowledge of mushroom diversity and distribution are imperative for successful conservation, management and optimum exploitation of the ecosystem for innumerable benefits to mankind (Nwordu et al. 2013). Mushrooms are important bioresource with diverse nutritional, medicinal and ecological benefits (Odeyemi et al. 2014). According to Chang and Miles (1992), and Das (2010), they are regarded as visible fungi with distinctive carpophores which represent the reproductive stage in the life cycles of Basidiomycetes and some Ascomycetes. Usually, mushroom carpophore gains more attention than its mycelia (vegetative stage) as a result of its sudden appearance in nature, characteristics morphology and color (Straatsma et al. 2001).

In the natural environment, mushrooms grow on variety of substrates, especially those containing lignin and cellulose, often abundant during the rainy season (Gbolagade 2005; Gbolagade et al. 2006; Jonathan et al. 2013). Their fleshy, spore-bearing fruiting bodies grow on soil or wood substrates whereas some exist in mycorrhizal relationship with trees (Onuoha and Obi-Adumanya 2010). Soil debris and dead woods are the most favorable environments for mushroom probably due to high content of degraded nutrients and capacity to retain moisture (Ayodele et al. 2011; Tibuhwa et al. 2011; Nwordu et al. 2013). Mushrooms have been reportedly found growing on

substrates like dead woods and base of dead woods, fallen trees, soil debris, palm trees and wastes, termite nests and cassava peels in forest and bush areas, grassy places, swamps, cocoa plantations, Fadama areas and burnt bushes (Gbolagade et al. 2006; Nwordu et al. 2013).

Globally, about 140,000 mushroom species are known to exist, with only about one-tenth taxonomically identified (Wasser 2002; Shelley and Geoffrey 2004). In Northern America, more than 70% in comparison with the globally identified mushrooms had been reported (CCB 2017). In India, about 10% had been investigated (Gurudevan et al. 2011).

Worldwide assessment of fungi diversity revealed that fungi distribution is mainly influenced by climates and certain types of habitat (Tedersoo et al. 2014); rainfall and temperature being the most important climatic elements (Straatsma et al. 2001). In contrast to what is obtainable in the temperate regions, mushrooms in the tropics are more diverse and are distributed over a smaller geographical area (Tedersoo et al. 2014). In Nigeria, mushrooms occur throughout the year (Ayodele et al. 2011) with the highest occurrence mostly in April to June (Nwordu et al. 2013). Onuoha and Obi-Adumanya (2010) stated that the boom of wild mushroom occurs once or twice a year and disappears within a month during the rainy season. However, desertification, deforestation, pesticides application, petroleum hydrocarbons and other environmental hazards contribute to the reduction in the quantities of mushroom

growing in the wild in many developing regions of the world (Okhuoya et al. 2010; Odeyemi et al. 2014).

Till date, the estimated mushrooms in the African continent is less than 1% of the total global estimate (Odebode 2005; Osemwegie and Okhuoya 2009; Oyetayo 2009; Bankole and Adekunle 2012; Nwordu et al. 2013). In most countries of the region, especially the most populated black nation Nigeria, there is paucity of information on the existing wild mushrooms. It is in the light of this that the present study investigated the ecology, diversity and seasonal distribution of wild mushrooms at ENPOST (Environmental Pollution Science and Technology) forest conservation.

MATERIALS AND METHODS

Description of the study area

The site of study for this research was the forest conservation section of ENPOST, located at Ido-Ijesa,

along Ife-Ilesa expressway, Ilesa, Southwestern Nigeria (Latitude 4°42'30"E to 4°42'45"E longitude 7°36'55"N to 7°37'10"N) (Figure 1). The forest conservation section of ENPOST has over 300 living species of trees. The predominant plant species in the forest is *Cleistopholis patens* (salt and oil tree). Others include *Elaeis guineensis* (palm tree), *Theobroma cacao* (cocoa tree) and *Cola nitida* (kolanut tree). Most trees within the forest are canopy-forming and some deciduous during the dry season. Green vegetations, dead wood logs, and tree debris such as fallen leaves and twigs which constitute soil debris are also abundant. Two different streams namely 'Aroni' and 'Eruru' flow along the forest making some areas marshy and muddy all year round. Interactions among fallen leaves and tree, moist soils, decayed matters, minimal inflow of anthropogenic activities and suitable environmental conditions make the forest suitable for the growth of different wild mushrooms.

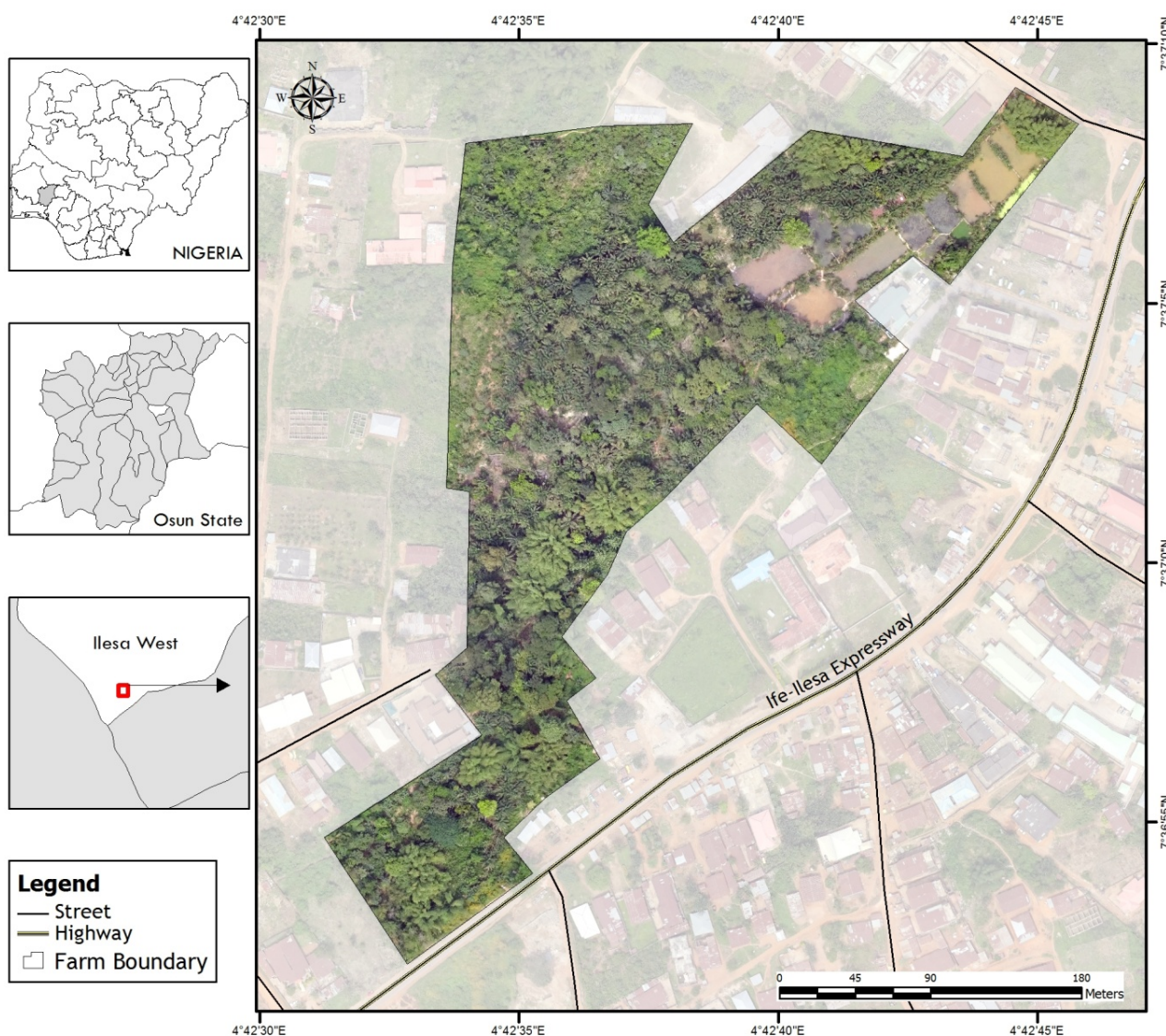


Figure 1. Map showing the location of ENPOST forest, Ilesa, Southwestern Nigeria

Presumptive identification of wild mushrooms

Specimens were initially photographed in their respective habitats and on growth substrates. Mushroom carpophores growing on soil and soil litter were carefully dug to prevent their bases from being bruised. Likewise, wood-rotting mushrooms were carefully detached from the bark of the trees. Representative mushroom specimens and wood substrates were collected in well-labeled sterile sampling paper bags and immediately taken to the laboratory for presumptive identification. Presumptive identification of mushroom samples was done using standard keys and by comparing the morphology of the carpophores with relevant literature (Rogers 1994; Nwordu et al. 2013). □

Ecology of wild mushrooms at ENPOST forest, Ilesa, Southern, Nigeria

Different indigenous mushrooms growing on the forest conservation section of ENPOST forest were studied over a 13-month regime (March 2014 and March 2015). Exactly 20 plots of 1000 m² were strategically mapped out on the site of study. Morphologically different wild mushrooms growing on different growth habitats and substrates within each plot were noted and counted three times a week. To avoid multiple counting of same mushroom carpophore, the mushrooms were marked with methylene blue at first encounter. The identity of wood substrates was done at the Herbarium unit of the Department of Pharmacognosy, Faculty of Pharmacy, Obafemi Awolowo University, Ile-Ife, Southwestern Nigeria.

Monitoring of climatic conditions of the site of study

The daily temperature and relative humidity of the study site were monitored using hygrometer (Fluke, 1620A, US), while daily rainfall data was obtained using rain gauge.

Data analysis

Diversity indices including species richness and diversity, relative abundance, dominance, and evenness were determined using BioDiversity Pro 2.0 software (McAleece et al. 1997). Species similarity between rainy and dry seasons was determined using the Jaccard's similarity coefficient (C_j). Mathematically, Jaccard coefficient (C_j) = $a / (a + b + c)$. Where a = number of mushroom species common to both rainy and dry seasons; b = number of mushroom species unique to rainy season; and c = number of mushroom species unique peculiar to dry season. Correlation was also performed to identify relationships among monthly climatic data, mushroom species richness and relative abundance using Statistical Package for Social Sciences [(SPSS) IBM version 22 software]. Statistical significance was set at $p < 0.05$.

RESULTS AND DISCUSSION

Enumeration of wild mushrooms found growing at ENPOST forest, Ilesa, Southwestern, Nigeria

During the period of this study, a total of 151 different mushroom species belonging to phyla Basidiomycota and

Ascomycota were found. Out of the 151 mushroom species, 143 (94.70%) belong to phylum Basidiomycota and 8 (5.30%) to phylum Ascomycota (Figure 2). Representative pictures are shown in Figure 3. □

All the mushroom samples surveyed belong to 68 genera and include: *Agaricus*, *Amanita*, *Anthrodiella*, *Auricularia*, *Cantharellula*, *Cantharellus*, *Clavatia*, *Clavariadelphus*, *Clitocybe*, *Clitopilus*, *Collybia*, *Coprinus*, *Coriolus*, *Cortinarius*, *Crepiotus*, *Cyathus*, *Dacrymyces*, *Daldinia*, *Dichometus*, *Dictophora*, *Ganoderma*, *Gleophyllum*, *Hapalopilus*, *Hericium*, *Heterobasidion*, *Hydnellum*, *Hydnum*, *Hygrocybe*, *Hypholoma*, *Hypoxylon*, *Inocybe*, *Inonotus*, *Laccaria*, *Lactarius*, *Lentinus*, *Lenzites*, *Lepiota*, *Leucocoprinus*, *Lycoperdon*, *Lyophyllum*, *Maramiellus*, *Marasmius*, *Micromphale*, *Mycena*, *Nidularia*, *Nolanea*, *Oxyporus*, *Panaeolina*, *Peziza*, *Phellinus*, *Piptoporus*, *Pleurotus*, *Pluteus*, *Polyporus*, *Psathyrella*, *Schizophyllum*, *Schizopora*, *Scleroderma*, *Stereum*, *Suillus*, *Termitomyces*, *Trametes*, *Trichoglossum*, *Tricholoma*, *Tricholomopsis*, *Tuber*, *Volvariella* and *Xylaria*.

Ecology of wild mushrooms at ENPOST forest, Ilesa, Southwestern, Nigeria

Mushrooms grew in two major habitats, i.e., ligneous (woody) and terrestrial (land). The species that grew in the ligneous habitats was 82 (54.31%) and higher than those on the terrestrial 68 (45.03%) (Figure 4). Interestingly, 1 (0.66%) species grew on termite nest (terrestrial) found on a living tangerine tree (ligneous) (Figure 5). All the mushroom species observed were habitat specific. □

The ligneous and terrestrial habitats comprise different growth substrates. While the terrestrial habitat consists of soil debris, termite mound and nest, bank of fish pond, soil heap, and sclerotium buried or on soil surface (Figure 6), it was root, stump, stem log, branch, leaves, and twig of dead *Cola nitida*, *Mangifera indica*, *Pseudospondias microcarpa*, *Elaeis guineensis*, *Triplochiton scleroxylon*, *Bambusa vulgaris*, *Cordyline australis*, *Gliricidia sepium*, *Musa paradisiaca*, *Pycnanthus angolensis* trees and some unidentified dead wood for the ligneous habitat (Table 1). All the mushrooms species from the terrestrial habitats were substrate specific with soil debris and heap having the highest 54 (79.41%) and lowest 1 (1.47%) number respectively (Figure 6). Of the 82 species that grew on the ligneous habitat, the highest number 59 (71.95%) grew on one type of growth substrate with Dead *Elaeis guineensis* supporting the highest number of species (25; 30.49%), 11 (13.41%) on two wood substrates simultaneously or at different times, 1 (1.22%) on 3 different substrates whereas 11 (13.41%) on unidentified substrates (Table 1). It was also observed that more than one mushroom species grew on some ligneous substrates at different times during this study. However, the succession pattern of mushrooms on each of the substrates was not studied. Notably were 3 mushroom species namely; *Hapalopilus nidulans*, *Lentinus subnudus* and *Pleurotus pulmonarius* that grew on erect dead trees (Unidentified tree, *Cola nitida*, *Cordyline australis* respectively) about 10 m above ground (Figure 7). All *Termitomyces* sp. both in the ligneous and terrestrial habitats were associated with termite mound or nest.

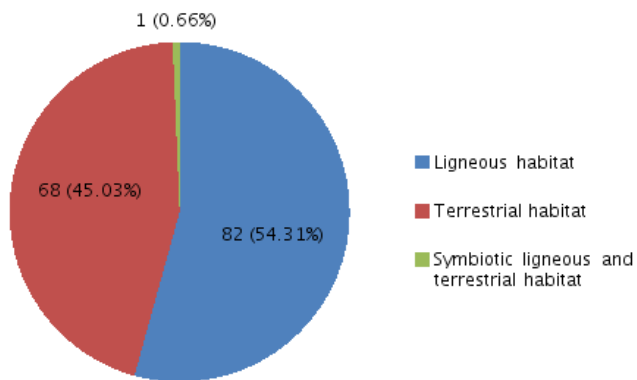


Figure 2. Percentage frequency of mushrooms in phyla Basidiomycota and Ascomycota

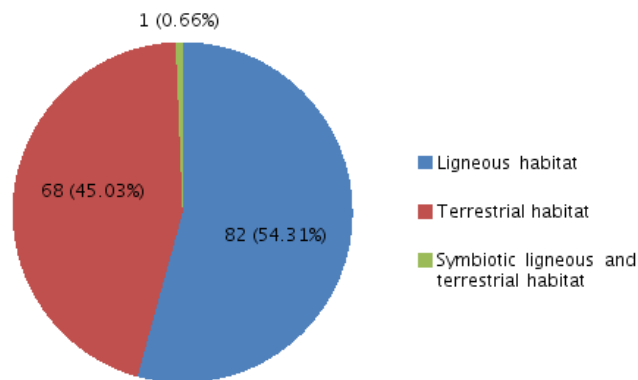


Figure 4. Distribution of mushroom species on different growth habitats at ENPOST forest, Ilesa, Southwestern Nigeria

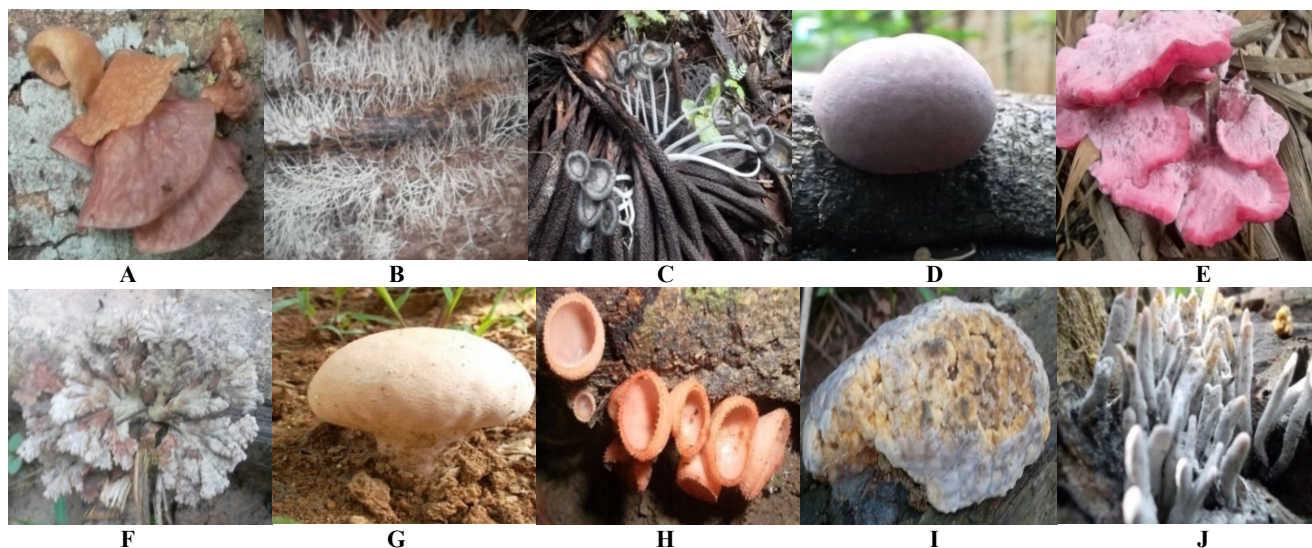


Figure 3. Representative pictures of wild mushrooms growing at ENPOST forest Ilesa, Southwestern, Nigeria. A. *Auricularia auricula-judae*; B. *Clavariadelphus junceus*; C. *Coprinus lagopides*; D. *Daldinia concentrica*; E. *Hydnellum peckii*; F. *Schizophyllum commune*; G. *Lycoperdon spadiceum*; H. *Peziza aurantia*; I. *Tuber excavatum*; J. *Xylaria hypoxylon*



Figure 5. A. *Stereum gausapatum* growing on termite nest on a living tangerine tree; B. Close-up view

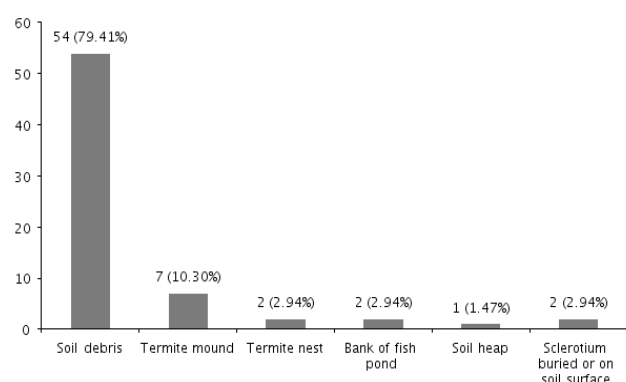


Figure 6. Frequency of mushroom species on the different terrestrial growth substrates

Monitoring of climatic conditions of the site of study

The lowest temperature (23°C) was obtained in August 2014 and the highest (28°C) in February 2015 (Table 2). The order of monthly precipitation was $16.85 > 12.55 > 9.09 > 6.58 > 5.66 > 2.46 > 2.39 > 2.11 > 0.41$ mm during May 2014, July 2015, October, August, September, June, April, March and November 2014 respectively. Only December 2014 and January 2015 had zero precipitation (Table 3). Based on the temperature and rainfall pattern of the site of study, March, April, November, December 2014 and January, February and March 2015 were grouped as dry season and May, June, July, August, September, and October 2014 as rainy season. Generally, the relative humidity was high throughout the study period. The highest (96.2 ± 2.2) and lowest (78.9 ± 3.2) values were recorded in July 2014 and March 2015 respectively (Table 2). □

Diversity indices of mushrooms sampled at the site

Mushroom species richness and relative abundance

Table 3 shows the monthly species richness of mushrooms at ENPOST forest between March 2014 and March 2015. Species richness was very high in October (70), followed by May (66) and September (62), 2014 and low March 2015 (11), January (13) and July (14) 2014.

Generally, mushrooms in the terrestrial habitat were abundant and found higher than those in the ligneous habitat only in May, September and October 2014. The relative abundance of mushroom carpophores in the terrestrial habitat during the period of study ranged from 0 to 45.35% with May 2014 recording the highest value (45.35%) (Table 3). In the ligneous habitat, the monthly relative abundance of mushroom carpophores ranged from 1.99% (August 2014) to 18.79% (April 2014) (Table 3). □

Mushroom dominance, species diversity and evenness

The dominance, species diversity, and evenness of the mushrooms studied in the rainy and dry seasons are presented in Table 4. Generally, a higher mushroom dominance was observed in the terrestrial habitat during the rainy and dry seasons (0.072 and 0.159 respectively) when compared to the ligneous habitat. On the contrary, species diversity was higher in both seasons in the ligneous habitat. Though the value obtained for evenness in the two seasons were higher in the terrestrial habitat with the highest evenness (0.581) during the dry season, no wide margin was noticed in both terrestrial and ligneous habitats during the rainy season where 0.261 and 0.264 were obtained respectively (Table 4). □

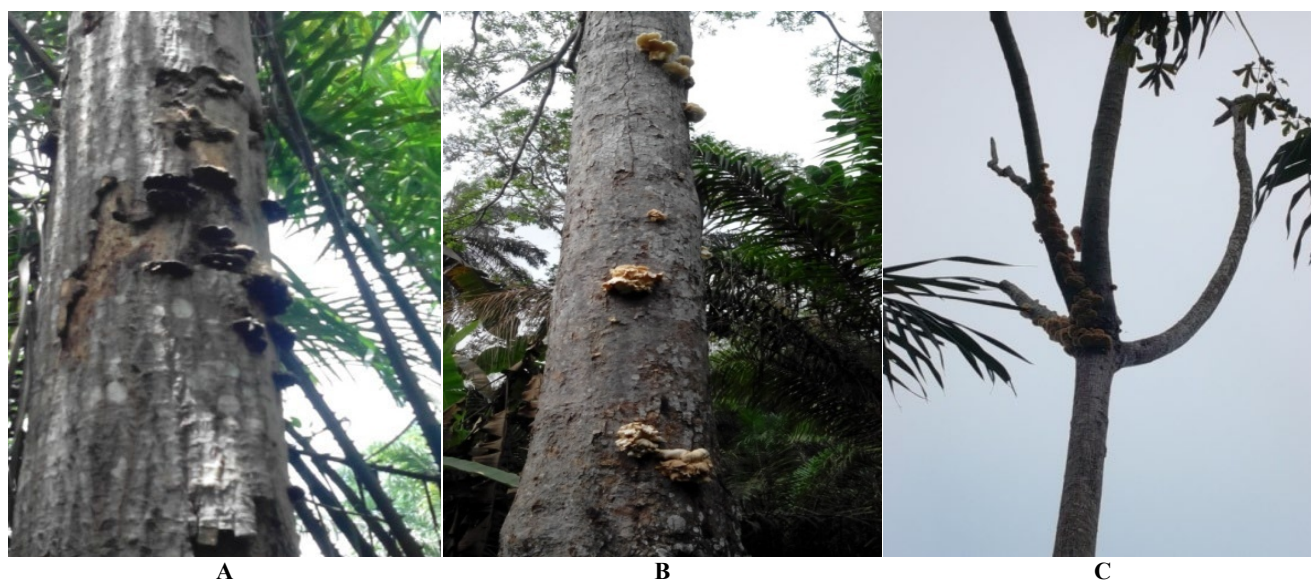


Figure 7. Pictures of mushrooms growing on erect dead wood logs about 10 m above ground. □ A. Unidentified tree, B. *Cola nitida*, C. *Cordyline australis*

Table 1. Occurrence and percentage frequency of mushroom species on different ligneous substrates

Substrate type	Substrate	n	%
Single substrate	Dead <i>Cola nitida</i>	14	17.07
	Dead <i>Elaeis guineensis</i>	25	30.49
	Dead <i>Mangifera indica</i>	13	15.85
	Dead burnt <i>Triplochiton scleroxylon</i> associated with termite nest	2	2.44
	Dead <i>Cordyline australis</i>	1	1.22
	Dead <i>Pseudospondias microcarpa</i>	1	1.22
	Dead <i>Bambusa vulgaris</i>	2	2.44
	Dead <i>Gliricidia sepium</i>	1	1.22
	Total	59	71.95
Dual substrates	Dead <i>Cola nitida</i> and Dead <i>Elaeis guineensis</i>	1	1.22
	Dead <i>Cola nitida</i> and Dead <i>Mangifera indica</i>	4	4.88
	Dead <i>Elaeis guineensis</i> and Dead <i>Musa paradisiaca</i>	2	2.44
	Living <i>Cola nitida</i> and <i>Elaeis guineensis</i>	1	1.22
	Dead <i>Elaeis guineensis</i> and Dead <i>Pycnanthus angolensis</i>	1	1.22
	Dead <i>Elaeis guineensis</i> and Dead <i>Bambusa vulgaris</i>	1	1.22
	Dead <i>Bambusa vulgaris</i> and Dead <i>Mangifera indica</i>	1	1.22
	Total	11	13.41
Triple substrates	Dead <i>Mangifera indica</i> , Dead <i>Elaeis guineensis</i> Font, Dead <i>Cordyline australis</i>	1	1.22
	Total	1	1.22
Unidentified substrates	Unidentified dead wood substrates	11	13.41
	Total	11	13.41
Overall total		82	100

Note: n - Number of mushroom species

Table 4. Seasonal variation in dominance, species diversity and evenness of mushrooms in the ligneous and terrestrial habitats during the rainy and dry season

Habitat	Rainy season			Dry season		
	Dominance	Species diversity	Evenness	Dominance	Species diversity	Evenness
Ligneous	0.033	3.912	0.261	0.093	3.019	0.220
Terrestrial	0.072	3.304	0.264	0.159	2.167	0.581

Table 2. Measurement of climatic elements of ENPOST forest, Ilesa, Southwestern Nigeria

Year	Month	Average monthly temp. (°C)	Number of rainfall (d) per month	Average monthly precip. (mm)	Average monthly relative humidity (%)
2014	March	27 ± 3	6	2.11	93.1 ± 2.7
	April	26 ± 3	6	2.39	92.7 ± 5.3
	May	26 ± 3	10	16.85	95.6 ± 2.8
	June	25 ± 3	9	2.46	94.9 ± 3.6
	July	24 ± 2	11	12.55	96.2 ± 2.2
	August	23 ± 3	8	6.58	92.8 ± 2.3
	September	24 ± 2	12	5.66	96.1 ± 2.6
	October	25 ± 2	13	9.09	95.8 ± 1.1
	November	26 ± 4	2	0.41	89.3 ± 2.7
	December	26 ± 5	0	0.00	85.3 ± 3.3
2015	January	26 ± 6	0	0.00	83.7 ± 4.5
	February	28 ± 6	ND	ND	80.2 ± 3.6
	March	27 ± 7	ND	ND	78.9 ± 3.2

Note: ND – Not determined

Table 3. Species richness and relative abundance of mushroom during the period of study

Year	Month	Species richness	Relative abundance of mushrooms in the terrestrial habitat (%)	Relative abundance of mushrooms in the ligneous habitat (%)
2014	March	20	0.32	7.57
	April	38	11.82	18.79
	May	66	45.35	8.91
	June	24	0.77	9.67
	July	14	0.00	2.11
	August	17	0.94	1.99
	September	62	12.22	10.28
	October	70	24.54	14.36
	November	42	2.68	13.75
	December	17	0.20*	3.27
2015	January	13	0.00	3.29
	February	26	0.45	3.57
	March	11	0.72	2.43
	Total		100	100

Note: *Mushrooms in this month were found growing close to the stream flowing through the forest

Seasonal variation in the occurrence and similarity of mushroom species found growing in the ligneous and terrestrial habitats

The occurrence of mushroom species was higher during the rainy season in the terrestrial habitat (n = 43) than in the ligneous habitat (n = 29). Contrariwise, the occurrence of mushroom species in the dry season was higher in the ligneous habitat (n = 14) than in the terrestrial habitat (n = 10). In addition, the number of mushroom species that occurred in both the dry and rainy seasons was higher in ligneous (39) than terrestrial habitats (15). Jaccard's similarity index indicates fewer mushroom species grew in both rainy and dry seasons in the terrestrial habitat when compared to the ligneous habitat (Table 5).

Generally in the ligneous habitat, the highest carpophores during the rainy and dry seasons was observed in *Coprinus disseminatus* (7,675) and *Schizophyllum commune* (7,329) respectively (Table 6) while it was *Mycena monticola* (760; rainy season) and *Mycena sanguinolenta* (362; dry season) in the terrestrial habitat (Table 7). In addition, *Auricularia auricula-judae*, *A. polytricha*, *Cantharellus cibarius*, *Coprinus lagopides*, *Cortinarius paleaceus*, *Inonotus hispidus*, *Lenzites betulina*, *Nidularia farcta*, *Pluteus salicinus*, *Schizophyllum commune*, *Trametes pubescens*, *Hericium abietis*, *Hypoxylon fragiforme*, and *Hypoxylon* sp. had more carpophores occurring in the dry season in the ligneous habitat (Table 6).

Table 5. Frequency and relationship between mushroom species found during the rainy and dry seasons

	Habitat	
	Ligneous	Terrestrial
Rainy season	29	43
Dry season	14	10
Both seasons	39	15
Jaccard's similarity index	0.4756	0.2206

Table 6. Occurrence of mushroom carpophores in the ligneous habitat

Mushroom identity	RS	DS	T
<i>Amanita bisporigera</i> □	300	269	569
<i>Amanita solitaria</i>	6	3	9
<i>Anthrodiella hoehnelii</i>	0	7	7
<i>Auricularia auricula-judae</i> □	1177	1843	3020
<i>Auricularia polytricha</i>	13	57	70
<i>Cantharellus cibarius</i>	78	137	215
<i>Clavariadelphus junceus</i>	1469	0	1469
<i>Clitocybe</i> sp.	44	0	44
<i>Coprinus disseminatus</i>	7675	498	8173
<i>Coprinus lagopides</i>	70	132	202
<i>Coprinus lagopus</i>	16	0	16
<i>Coprinus plicatilis</i>	0	4	4
<i>Coprinus</i> sp.1	0	6	6
<i>Cortinarius paleaceus</i>	1452	1629	3081
<i>Crepiotus mollis</i>	0	14	14
<i>Cyathus striatus</i>	0	26	26
<i>Dacrymyces</i> sp.	586	0	586

<i>Daldina concentrica</i>	16	16	32
<i>Dichomitus campestris</i>	3	0	3
<i>Ganoderma applanatum</i>	6	3	9
<i>Ganoderma resinaceum</i>	39	29	68
<i>Gleophyllum sepium</i>	0	1	1
<i>Hapalopilus nidulans</i>	0	158	158
<i>Hericium abietis</i>	78	121	199
<i>Heterobasidion annosum</i>	2	2	4
<i>Hydnellum peckii</i>	71	6	77
<i>Hydnum</i> sp.	137	80	217
<i>Hypoxylon fragiforme</i> □	23	38	61
<i>Hypoxylon</i> sp.□	28	44	72
<i>Inonotus hispidus</i>	7	31	38
<i>Inonotus radiatus</i>	45	32	77
<i>Lactarius obscuratus</i>	9	0	9
<i>Lentinus subnudus</i>	1011	407	1418
<i>Lenzites betulina</i>	119	210	329
<i>Lepiota ventriosospora</i>	24	0	24
<i>Lycoperdon echinatum</i>	15	0	15
<i>Lyophyllum loricatum</i>	6	0	6
<i>Marasmiellus candidus</i>	2081	2910	4991
<i>Marasmius</i> sp.1	22	0	22
<i>Marasmius</i> sp.2	0	67	67
<i>Marasmius wynnei</i>	12	0	12
<i>Marasmius ramealis</i>	1099	749	1848
<i>Mycena galericulata</i>	1	0	1
<i>Mycena inclinata</i>	426	0	426
<i>Mycena maculata</i>	0	137	137
<i>Mycena</i> sp.1	32	0	32
<i>Mycena</i> sp.2	279	2	281
<i>Mycena tortuosa</i>	13	0	13
<i>Mycena vitilis</i>	1421	721	2142
<i>Nidularia farcta</i> □	67	78	145
<i>Nolanea cetrata</i>	168	0	168
<i>Oxyporus populinus</i>	26	0	26
<i>Panaeolus foenisecii</i>	23	0	23
<i>Panaeolus semiovatus</i>	9	0	9
<i>Peziza aurantia</i>	290	51	341
<i>Phellinus igniarius</i>	2	0	2
<i>Piptoporus betulinus</i>	1	0	1
<i>Pleurotus florida</i>	27	9	36
<i>Pleurotus lignatilis</i>	129	100	229
<i>Pleurotus ostreatus</i>	3	0	3
<i>Pleurotus pulmonarius</i>	0	78	78
<i>Pluteus salicinus</i>	1	5	6
<i>Polyporus floccipes</i>	3	0	3
<i>Polyporus</i> sp.	231	0	231
<i>Polyporus varius</i>	6	0	6
<i>Psathyrella marcescibilis</i>	0	1	1
<i>Schizophyllum commune</i>	3408	7329	10737
<i>Schizopora paradoxa</i>	15	33	48
<i>Scleroderma areolatum</i>	10	0	10
<i>Sterum complicatum</i>	22	20	42
<i>Sterum hirsutum</i>	9	20	29
<i>Stereum</i> sp.	3	0	3
<i>Termitomyces</i> sp.	0	2	2
<i>Termitomyces striatus</i>	0	3	3
<i>Trametes ochracea</i>	274	146	420
<i>Trametes pubescens</i>	18	20	38
<i>Trametes versicolor</i>	56	47	103
<i>Trichoglossum hirsutum</i>	41	0	41
<i>Tricholomopsis</i> sp.	0	7	7
<i>Tuber excavatum</i>	5	0	5
<i>Xylaria hypoxylon</i>	1515	1178	2693
<i>Xylaria longipes</i>	3036	2372	5408
Total	29309	21888	51197

Note: Legend: RS – Rainy season; DS – Dry season; T – Total

Table 7. Occurrence of mushroom carpophores in the terrestrial habitat

Mushroom identity	RS	DS	T
<i>Agaricus impudicus</i>	0	20	20
<i>Agaricus semotus</i>	55	0	55
<i>Agaricus silvicola</i>	35	0	35
<i>Amanita cokeri</i>	4	17	21
<i>Amanita muscaria</i>	1	0	1
<i>Amanita parvicolvata</i>	0	6	6
<i>Amanita phalloides</i>	20	11	31
<i>Amanita virosa</i>	1	0	1
<i>Cantharellula umbonata</i>	6	31	37
<i>Clavatia excipuliformis</i>	1	1	2
<i>Clitocybe nud</i>	2	0	2
<i>Clitopilus prunulus</i>	6	0	6
<i>Collybia distorta</i>	0	9	9
<i>Collybia erythropus</i>	24	0	24
<i>Collybia maculate</i>	16	0	16
<i>Collybia palustris</i>	0	6	6
<i>Collybia</i> sp.	51	0	51
<i>Coprinus palustris</i>	0	1	1
<i>Coprinus</i> sp.2	24	22	46
<i>Coprinus</i> sp.3	1	1	2
<i>Dictophora duplicata</i>	8	0	8
<i>Hygrocybe cantherelus</i>	235	0	235
<i>Hygrocybe conica</i>	1	0	1
<i>Hygrocybe miniata</i>	31	0	31
<i>Hygrocybe nigrescens</i>	2	0	2
<i>Hygrocybe vitellina</i>	485	0	485
<i>Hypholoma capnoides</i>	17	0	17
<i>Inocybe fastigiata</i>	2	5	7
<i>Laccaria laccata</i>	375	0	375
<i>Lactarius deliciosus</i>	1	0	1
<i>Lepiota adulterine</i>	4	0	4
<i>Lepiota felina</i>	13	0	13
<i>Lepiota konrandi</i>	1	0	1
<i>Lepiota mastoidea</i>	13	0	13
<i>Lepiota pseudohelveola</i>	10	0	10
<i>Leucocoprinus birnbaumii</i>	2	0	2
<i>Leucocoprinus brebissonii</i>	0	5	5
<i>Lycoperdon spadiceum</i>	1	0	1
<i>Macrolepiota procera</i>	56	14	70
<i>Marasmius epiphyllus</i>	32	0	32
<i>Marasmius</i> sp.3	24	0	24
<i>Marasmius</i> sp.4	33	0	33
<i>Marasmius</i> sp.5	125	0	125
<i>Micromphale brassicolens</i>	0	2	2
<i>Micromphale foetidum</i>	420	0	420
<i>Mycena acicula</i>	3	0	3
<i>Mycena aetitis</i>	3	0	3
<i>Mycena alcalina</i>	1	0	1
<i>Mycena crocata</i>	12	21	33
<i>Mycena galopus</i>	78	9	87
<i>Mycena haematopus</i>	67	0	67
<i>Mycena interrupta</i>	69	0	69
<i>Mycena monticola</i>	760	0	760
<i>Mycena pearsoniana</i>	43	0	43
<i>Mycena sanguinolenta</i>	0	362	362
<i>Panaeolina foenisecii</i>	11	3	14
<i>Pleurotus tuber-regium</i>	20	8	28
<i>Suillus</i> sp.	16	0	16
<i>Termitomyces aurantiacus</i>	442	0	442
<i>Termitomyces bulborhizus</i>	4	0	4
<i>Termitomyces letestui</i>	202	0	202
<i>Tricholoma inocybeoides</i>	84	34	118
<i>Tricholoma</i> sp.	0	1	1
<i>Tricholoma sulphureum</i>	0	1	1
<i>Tricholoma ustale</i>	82	49	131
<i>Tricholoma vaccinum</i>	3	8	11
<i>Tricholomopsis decora</i>	3	0	3
<i>Volvariella speciosa</i>	1	0	1
Total	4042	647	4689

Note: Legend: RS – Rainy season; DS – Dry season; T – Total

Likewise, in the terrestrial habitat, *Amanita cokeri*, *Cantharellula umbonata*, *Mycena crocata* and *Tricholoma vaccinum* had more carpophores in the dry season (Table 7). Whereas *Schizophyllum commune* recorded the highest carpophores in the ligneous habitat (10,737), it was *Mycena monticola* in the terrestrial habitat (760) (Table 6-7). Overall, the total frequency of mushroom carpophores was generally higher in the rainy than dry season in both habitats (Table 6-7).

Correlations among monthly climatic data, mushroom species richness and relative abundance

The result obtained shows strong significant relationship among species richness, relative abundance of mushrooms in the terrestrial ($r = 0.829$; $p = 0.000$) and ligneous ($r = 0.679$; $p = 0.011$) habitat. Average monthly precipitation correlated strongly with relative abundance of mushroom carpophores in the terrestrial habitat ($r = 0.7018$, $p = 0.024$) and number of rainfall per month ($r = 0.730$; $p = 0.011$). Also, average monthly relative humidity correlated strongly with average monthly temperature ($r = -0.668$; $p = 0.013$), number of rainfall per month ($r = 0.952$; $p = 0.000$) and average monthly precipitation ($r = 0.689$; $p = 0.019$) (Table 8).

Discussion

Forest structure including degree of tree cover, humidification of soil, nature of substratum, acidity of soil and favorable environmental condition are pivotal to the growth, development and diversity of mushrooms in the biological world (Straatsma et al. 2001; Odeyemi et al. 2014; Djelloul and Samraoui, 2011; Johnsy et al. 2011). In the present study, the 151 mushrooms identified have been widely reported in India (Dwivedi et al. 2012; Pushpa and Purushothama 2012; Vyas et al. 2014), Bangladesh (Rashid et al. 2016; Romainul and Aminuzzaman 2016), Cameroon (Andrew et al. 2013), Algeria (Djelloul and Samraoui 2011), Switzerland (Straatsma et al. 2001) and Mexico (LaRochelle and Berkes 2003).

In Nigeria, Zoberi (1973), Oso (1975, 1977a, b), Alofe (1985), Gbolagade et al. (2006), Ezeibekwu et al. (2009), Oyetayo (2009, 2011), Okhuoya et al. (2010), Adedayo (2011), Ayodele et al. (2011), Nwordu et al. (2013) had earlier reported mushrooms in the genera *Agaricus*, *Amanita*, *Auricularia*, *Clavatia*, *Collybia*, *Coprinus*, *Daldina*, *Dictophora*, *Ganoderma*, *Lactarius*, *Lentinus*, *Lepiota*, *Lycoperdon*, *Pleurotus*, *Psathyrella*, *Schizophyllum*, *Termitomyces*, *Tricholoma*, *Volvariella* and *Xylaria*. This is the first time mushrooms in the genera *Anthrodiella*, *Cantharellula*, *Cantharellus*, *Clavaria-delphus*, *Clitocybe*, *Clitopilus*, *Coriolus*, *Cortinarius*, *Crepitatus*, *Cyathus*, *Dacrymyces*, *Dichometus*, *Gleophyllum*, *Hapalopilus*, *Hericius*, *Heterobasidion*, *Hydnellum*, *Hydnum*, *Hygrocybe*, *Hypholoma*, *Hypoxylon*, *Inocybe*, *Inonotus*, *Laccaria*, *Lenzites*, *Leucocoprinus*, *Lyophyllum*, *Maramiellus*, *Marasmius*, *Micromphale*, *Mycena*, *Nidularia*, *Nolanea*, *Oxyporus*, *Panaeolina*, *Peziza*, *Phellinus*, *Piptoporus*, *Pluteus*, *Polyporus*, *Schizopora*, *Scleroderma*, *Stereum*, *Suillus*, *Trametes*, *Trichoglossum*, *Tricholomopsis* and *Tuber* are reported.

Table 8. Correlations among monthly climatic data, mushroom species richness and relative abundance

		1	2	3	4	5	6	7
1	Species richness	1						
2	Relative abundance of mushrooms in the terrestrial habitat (%)	0.829** (0.000)	1					
3	Relative abundance of mushrooms in the ligneous habitat (%)	0.679* (0.011)	0.434 (0.138)	1				
4	Average monthly temperature	-0.148 (0.630)	-0.054 (0.861)	0.009 (0.977)	1			
5	Number of rainfall (d) per month	0.555 (0.076)	0.479 (0.136)	0.205 (0.546)	-0.542 (0.085)	1		
6	Average monthly precipitation (mm)	0.466 (0.148)	0.716* (0.013)	-0.089 (0.794)	-0.353 (0.287)	0.730* (0.011)	1	
7	Average monthly relative humidity (%)	0.534 (0.060)	0.453 (0.120)	0.467 (0.107)	-0.668* (0.013)	0.952** (0.000)	0.689* (0.019)	1

Note: Values in parenthesis are p-value of correlation coefficient □

Majority of wild mushrooms are found on variety of organic substrates in ligneous and terrestrial habitats where they obtain nutrients for growth (Gbolagade et al. 2006; Johnsy et al. 2011, Nwordu et al. 2013). In the current study, a high occurrence of wild mushrooms was recorded in the ligneous compared to terrestrial habitat and their symbiotic association (Figure 4). This could be attributed to the abundance of woody substrates and their colonization with mushroom mycelia that are mostly saprophytic. Mushrooms can secrete hydrolytic and oxidative extracellular enzymes that digest lignin and cellulose present in the substrates into smaller organic compounds or minerals into usable forms (Altaf et al. 2010; Odeyemi et al. 2014). Similarly, the high prevalence of terrestrial mushrooms on soil debris (79.41%) (Figure 6) indicates presence of simple organic matter and massive mycelia colonization of the underground soil. Mushroom develop from undisturbed interwoven web of hyphae called mycelium, which often time is buried in or found on the substrate where it derives its nourishment (Wilkinson and Buezaeki 1982; Ramsbottom 1989; Adedayo 2011). Also, association of *Termitomyces* species with termite mound or nest was noted by other researchers (Gbolagade 2006; Johnsy et al. 2011).

Previous investigations have documented association of some mushrooms with a particular genus or family of trees (Arora 2008; Karwa and Rai 2010), suggesting mushroom substrate specificity. In this study, quite a high percentage of the observed mushrooms (about 72%) were associated with single substrate and about 15% with two or three substrates (Table 1) revealing that all mushrooms are not substrate-specific.

Spore dispersal is crucial for sustainability of mushroom in nature. During the reproductive stage in the life cycle of mushrooms, spores are released from blown out mushroom carpophores and settle on substrates where it establishes its mycelium and continues the cycle (Odeyemi et al. 2014). The growth of mushrooms on erect dead trees

5 feet above ground (Figure 7) revealed that some mushroom spores might be carried up in the air by current, settles on and colonize suitable wood substrate which can support their growth. Flying insects, pecking birds and crawling animals carrying mushroom spores might also be agent of dispersal (Abbott 2002; Lilleskov and Bruns 2005; Viljanen-Rollinson et al. 2007).

The ranges of temperature (from 23 to 28°C), precipitation (from 0.00 to 16.85 mm) and high relative humidity (from 78.9 to 96.1%) (Table 2) observed in this study fell within the established limits that support the growth of mushrooms (Odeyemi et al. 2014). However, the temperature and rainfall range slightly differ compared to the report of Straatsma et al. (2001) on the biodiversity of fungal carpophore in a Swiss forest plot over 21 years regime. They reported a temperature range of 16.1 to 20.6°C yearly and a range of 285.5 to 636.3 mm precipitation yearly on an average of 6 to 10 months. The occurrence of mushrooms in all the months of study showed their ready availability all-year-round at ENPOST forest. This is in agreement with the report of Adedayo (2011) who also noted the growth of mushrooms throughout the year. □

Mushroom diversity expressed as species richness, relative abundance in ligneous and terrestrial habitats reveals the dynamic nature of mushrooms between months, habitats and seasons. Species richness peaked in May (66), September (62) and October (70) 2014 with corresponding high mushroom relative abundance, 45.35, 12.22 and 24.54%, in the terrestrial habitat, compared to the ligneous counterpart (Table 3). The boom of mushroom in May align with the report of Nwordu et al. (2013) who stated that highest occurrence of mushroom falls between April and June. Other diversity indices including dominance, species diversity, evenness and Jaccard's index give insight into preponderance, degree of representation and possible overlap of species and environmental stability in the forest (Kindt and Coe 2005). Dominance index indicated that

more species dominated in the terrestrial habitat comparable to the ligneous habitat. The high species diversity index in ligneous (3.912) and terrestrial (3.304) habitats (Table 4) infer a more stable and less disturbed environment during rainy season; albeit, the community was more stable. Similarly, the high evenness index in the terrestrial habitat (0.581) indicated that species occurring during the dry season are common and abundant. Jaccard's index also revealed that more species of mushrooms occurred both in the rainy and dry seasons in ligneous habitat than the terrestrial. □

A diverse mushroom community observed at ENPOST forest may not be a true representation of all the mushrooms present as some mycelia remain dormant underground or in their substrate until there is a prevailing environmental condition favorable for its growth (Adedayo, 2011; Andrew et al. 2013). Adequate moisture and water holding capacity of growth substrate are key factors to be considered for mushroom development (Ayodele et al. 2011; Tibuhwa et al. 2011; Bellettini et al. 2016). In this study, mushrooms found during rainy season were higher than dry season (Table 5). In the same vein, frequencies of carpophore obtained suggest that moisture requirement for the mushrooms differ, i.e., species can grow either under high, low moisture availability or both (Tables 6, 7). The present findings also reveal that monthly precipitation correlated significantly with relative abundance of mushrooms in the terrestrial habitat.

In conclusion, different mushroom species are still relatively abundant in Nigeria due to prevailing environmental and climatic conditions and copious availability of growth substrates which support their growth. The ecology, biodiversity and seasonal distribution of wild mushrooms at ENPOST forest were studied. Our study revealed for the first time in Nigeria a total of one hundred and fifty-one different mushrooms, a good representative of Nigeria's rich mushroom flora, which can be of diverse human and biotechnological benefits if properly harnessed. However, it should be noted that these bioresources are gradually going into extinction as a result of human activities such as deforestation, bush burning, use of pesticides and herbicides, urbanization and climate change. Hence, need for their urgent preservation and sustainability is advocated.

ACKNOWLEDGEMENTS

We sincerely appreciate the Management of ENPOST Forest, Ilesa, Nigeria for granting access to the site of study.

REFERENCES

- Abbott SP. 2002. Insects and other arthropods as agents of vector-dispersal in fungi. www.thermapure.com/pdf/AbbottInsectdispersal.pdf
- Adedayo MR. 2011. Proximate analysis on four edible mushrooms. *J Appl Sci Environ Manag* 15 (1): 9-11.
- Altat SA, Umar DM, Muhammad MS. 2010. Production of xylanase enzyme by *Pleurotus eryngii* and *Flammulina velutipes* grown on different carbon sources under submerged fermentation. *World Appl Sci J* 8: 47-49.
- Alofe FV. 1985. The General Characteristics and Cultivation of Some Nigerian Mushrooms. [Dissertation]. Obafemi Awolowo University, Ile-Ife. [Nigeria]
- Andrew EA, Kinge TR, Tabi EM, Thiobal N, Mih AF. 2013. Diversity and distribution of macrofungi (mushrooms) in the Mount Cameroon Region. *J Ecol Nat Environ* 5 (10): 318-334.
- Arora D. 2008. Notes on economic mushrooms: Xiao Ren Ren: the little people of Yunnan. *Econ Bot* 62: 541-544.
- Ayodele SM, Akpaja EO, Adamu Y. 2011. Some edible medicinal mushrooms of Igala land in Nigeria, their sociocultural and ethnomycological uses. *Int J Sci Nat* 2 (3): 473-476.
- Bankole PO, Adegkunle AA. 2012. Studies on biodiversity of some mushrooms collected in Lagos State, Nigeria using biotechnological methods. *J Yeast Fungal Res* 3 (4): 37-48.
- Bellettini MB, Fiorda FA, Maievas HA, Teixeira GL, Avila S, Homung PS, Júnior AM, Ribani RH. 2016. Factors affecting *Pleurotus* spp. *Saudi J Biol Sci*. DOI: 10.1016/j.sjbs.2016.12.005.
- Chang ST, Miles PG. 1992. Mushroom biology – a new discipline. *The Mycologist* 6: 64-65.
- Cofrin Center for Biodiversity (CCB). 2017. Biodiversity of macrofungi in Northern Door Country, WI. University of Wisconsin, Green-Bay, WI.
- Das K. 2010. Diversity and conservation of wild mushrooms in Sikkim with special reference to Barsey Rhododendron Sanctuary. *NeBio* 1: 1-13.
- Djelloul R, Samraoui B. 2011. Distribution and ecology of the superior mushrooms of the Aulnaie of Ain Khair (El Kala National Park, Northeastern Algeria). *Afr J Environ Sci Technol* 5 (6): 448-456.
- Dwivedi S, Tiwari MK, Chauhan UK, Pandey AK. 2012. Biodiversity of mushrooms of Amarkantak biosphere reserve forest of Central India. *Int J Pharm Life Sci* 3 (1): 1363-1367.
- Ezeibekwe IO, Ogbonnaya CI, Unamba CIN, Osuala OM. 2009. Proximate analysis and mineral composition of edible mushrooms in parts of SouthEastern Nigeria. *Report and Opinion* 1 (4): 32-36.
- Gbolagade JS. 2005. Bacteria associated with cultures of *Psathyrella atrombonata* (Pegler), and *Schizophyllum commune* (Fr.Ex.Fr), Nigerian edible mushrooms. *Acta Phytopathologica. Et Entomologica Hungarica* 40 (2-3): 333-340.
- Gbolagade J, Ajayi A, Oku I, Wankasi D. 2006. Nutritive value of common wild edible mushrooms from Southern Nigeria. *Global J Biotechnol Biochem* 1 (1): 16-21.
- Gurudevan T, Prakasam V, Chandrasekar G, Sakthivel K, Veeralakshmi S, Velazhahan R, Kalaiselvi G. 2011. Biodiversity, conservation, and utilization of mushroom flora from the Western Ghats region of India. In: *Proceedings of the 7th International Conference on Mushroom Biology and Mushroom Products (ICMBMP7)*. Arcachon, 4-7 October 2011. [France]
- Johnsy G, Sargunam SD, Dinesh MG, Kaviyaran V. 2011. Nutritive value of edible wild mushrooms collected from the Western Ghats of Kanyakumari District. *Botany Research International* 4 (4): 69-74.
- Jonathan SG, Nwokolo VM, Ekpo EN. 2013. Yield performance of *Pleurotus pulmonarius* (Fries.) Quelet, cultivated on different agro-forest wastes in Nigeria. *World Rural Observ* 5 (1): 22-30. □
- Karwa ALKA, Rai MK. 2010. Tapping into the edible fungi biodiversity of Central India. *Biodiversitas* 11 (2): 97-101.
- Kindt R, Coe R. 2005. Tree diversity analysis. A manual and software for common statistical methods for ecological and biodiversity studies. [Nairobi] □
- LaRochelle S, Berkes F. 2003. Traditional ecological knowledge and practice for edible wild plants: biodiversity use by the Raramuri in the Sierra Tarahumara, Mexico. *Int J Sust Dev World Ecol* 10: 361-375.
- Lilleskov EA, Bruns TD. 2005. Spore dispersal of a resupinate ectomycorrhizal fungus, *Tomentella subulacina*, via soil food webs. *Mycologia* 97 (4): 762-769.
- McAleece N, Gage JDG, Lambshead PJO, Paterson GLJ. 1997. BioDiversity professional statistics analysis software. Jointly developed by the Scottish Association for Marine Science and the Natural History Museum, London. □
- Nwordu ME, Isu RN, Ogbadu GH. 2013. Catalogue and identification of some wild edible macro-fungi in Nigeria. *Int J Food Sci* 2 (1): 1-15.
- Odebo SO. 2005. Contributions of selected non-timber forest products to household food security in Nigeria. *J Food Agric Environ* 3: 138-141.

- Odeyemi O, Adeniyi MA, Odeyemi Y. 2014. Introduction to Tropical Mycology. Universal Academic Press, China.
- Okhuoya JA, Akpaja EO, Osemwegie OO, Oghenekaro AO, Ihayere CA. 2010. Nigeria mushrooms: underutilized non-wood forest resources. *J Appl Sci Environ Manag* 14 (1): 43-54.
- Onuoha CI, Obi-Adumanya GA. 2010. Proximate analysis of *Pleurotus tuber-regium* (Sing) grown on the different substrates. *Researcher* 2 (10): 7-11.
- Osemwegie OO, Okhuoya JA. 2009. Diversity of macrofungi in oil palm agroforests of Edo State, Nigeria. *J Biol Sci* 9 (6): 584-593.
- Oso BA. 1975. Mushrooms and the Yoruba people of Nigeria. *Mycologia* 67: 311-319.
- Oso BA. 1977a. *Pleurotus tuber-regium* from Nigeria. *Mycologia* 69: 271-279.
- Oso BA. 1977b. Mushroom in Yoruba mythology and medicinal practices. *Econ Bot* 31: 367-371.
- Oyetayo OV. 2011. Medicinal uses of mushrooms in Nigeria: Towards full and sustainable exploitation. *Afr J Tradit Complement Altern Med* 8 (3): 267-274.
- Oyetayo VO. 2009. Molecular characterisation of *Termitomyces* species collected from Ado Ekiti and Akure, Nigeria. *Nig J Microbiol* 23 (1): 1933-1938.
- Pushpa H, Purushothama KB. 2012. Biodiversity of mushrooms in and around Bangalore (Karnataka), India. *Am Eurasian J Agric Environ Sci* 12 (6): 750-759.
- Ramsbottom J. 1989. Mushrooms and Toadstools. Bloomsbury Books, London.
- Rashid SN, Aminuzzaman FM, Islam MR, Rahaman M, Romainul MI. 2016. Biodiversity and distribution of wild mushrooms in the Southern Region of Bangladesh. *J Adv Biol Biotechnol* 9 (1): 1-25.
- Rogers P. 1994. Mushrooms and other Fungi of Great Britain and Europe. Macmillan Publishers Limited, London.
- Romainul MI, Aminuzzaman FM. 2016. Macrofungi biodiversity at the Central and Northern biosphere reserved areas of tropical moist deciduous forest region of Bangladesh. *J Agric Ecol Res Intl* 5 (4): 1-11.
- Shelley E, Geoffrey K. 2004. Pocket nature: Fungi. Dorling Shelley Evans and Geoffrey Kindersley Limited, London.
- Straatma G, Ayer F, Nd Egli S. 2001. Species richness, abundance and phenology of fungal fruitbodies over 21 years in a Swiss forest plot. *Mycol Res* 105 (5): 525-523.
- Tedersoo L, Bahram M, Põlme S, Kõljalg U, Yorou NS, Wijesundera R, Villarreal RL, Vasco-Palacios AM, Thu PQ, Suija A, Smith ME, Sharp C, Saluveer E, Saitta A, Rosas M, Riit T, Ratkowsky D, Pritsch K, Põldmaa K, Piepenbring M, Phosri C, Peterson M, Parts K, Pärtel K, Otsing E, Nouhra E, Njouonkou AL, Nilsson RH, Morgado LN, Mayor J, May TW, Majuakim L, Lodge DJ, Lee SS, Larsson KH, Kohout P, Hosaka K, Hiiesalu I, Henkel TW, Harend H, Guo LD, Greslebin A, Grelet G, Geml J, Gates G, Dunstan W, Dunk C, Drenkhan R, Dearnaley J, De Kesel A, Dang T, Chen X, Buegger F, Brearley FQ, Bonito G, Anslan S, Abell S, Abarenkov K. 2014. Global diversity and geography of soil fungi. *Science* 346 (6213): 1256688.
- Tibuhwa DD, Muchane MN, Masiga CW, Mugoya C, Muchai M. 2011. An inventory of macro-fungi and their diversity in the Serengeti-Masai Mara ecosystem, Tanzania and Kenya. *J Biol Sci* 11 (6): 399-410.
- Viljanen-Rollinson SLH, Parr EL, Marroni MV. 2007. Monitoring long-distance spore dispersal by wind - a review. *N Z Plant Prot* 60: 291-296.
- Vyas D, Chaubey A, Dehariya P. 2014. Biodiversity of mushrooms in Patharia forest of Sagar (MP)-III. *Int J Biodivers Conserv* 6 (8): 600-607.
- Wasser S. 2002. Medicinal mushrooms as a source of anti-tumor and immune-modulating polysaccharides. *Appl Microbiol Biotechnol* 60: 258-274.
- Wilkinson J, Buezaeki S. 1982. Mushrooms and Toadstools. Harper Collins, Glasgow.
- Zoberi MH. 1973. Some edible mushrooms from Nigeria. *Nig Field* 38: 81-90.