

Rhizosphere fungal community, soil physicochemical properties, understorey vegetation and their relationship during post-coal mining reclamation in East Kalimantan, Indonesia

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Abstract. Sudrajat, Widhayasa B, Rusdiansyah, Susanto D. 2019. Rhizosphere fungal community, soil physicochemical properties, understorey vegetation and their relationship during post-coal mining reclamation in East Kalimantan, Indonesia. *Biodiversitas* 20: 1953-1962. The objective of this study was to evaluate the composition of fungal community in rhizosphere, understorey vegetation and soil physicochemical properties post-coal mining reclamation. Soil sampling was conducted using a series of reclamation stages plots (undisturbed, disturbed, two, nine, twelve and sixteen years of reclamation) with drill ground at a depth of 0-30 cm. On each plot, five-point drilling was conducted; soil samples were composited; and their physicochemical properties including soil pH, organic carbon, total N, available P, available K, Ca, Mg, Fe, Mn, and S content were analyzed. Rhizosphere microfungi were isolated by using serial dilution and plated on potato dextrose agar added with Streptomycin. Identification and characterization of soil microfungi were done with the help of accessible manuals of fungi. The results showed that fungal genera at rhizosphere during post-coal mining reclamation were *Aspergillus*, *Penicillium*, *Cephalosporium*, *Gliocladium*, *Humicola*, *Trichoderma*, and *Paecilomyces*. *Aspergillus* was dominant comprising 74% of the total isolates. The fungal density in the undisturbed soil and the post-coal mining reclamation soil were higher than in the disturbed soil. The fungal rhizosphere groups isolated were saprophytic fungi that were present in a high relative density. In addition, the results showed that fungal rhizosphere was negatively correlated with soil pH and organic carbon. On the other hand, fungal rhizosphere was positively correlated with total N, available P, and available K. It can be concluded that the time period of post-coal mining reclamation, vegetation, and soil physicochemical profoundly determined the soil fungal community.

Keywords: Microfungal rhizosphere, post-coal mining reclamation, soil physicochemical

INTRODUCTION

The activities at coal mining sites cause drastic disturbances in a landscape and alter the ecological environment, thereby disrupting the functional stability of the soil microbial community. This condition needs to be rehabilitated by conducting reclamation and revegetation. The ultimate goal of mine land reclamation and revegetation is the reestablishment of a productive, healthy, and sustainable ecosystem that is suitable for land use. Currently, only aboveground indicators such as soil erosion and vegetation characteristics such as production, cover, diversity, and shrub density are considered in determining reclamation success.

The long-term goal of reclamation and revegetation is to improve the condition of the microclimate, biodiversity, wildlife habitats, and the condition of the land to be more protected, conserved and productive (Mummey et al. 2002; Rana et al. 2007; Yusuf and Arisoelaningsih 2017). The most important aspect in ecosystem recovery is soil characteristic. Soil quality serves so many important ecosystems for not only human beings but also the other organism in an environment (Noviyanto et al. 2017). The

microbial community in soil is fundamental for organic matter transformation and plays a principal role in the biogeochemical cycles in this ecosystem. Soil microbes are vital for the continuous cycling of nutrient and for driving aboveground ecosystems. Considering its importance, restoration of soil structure and function is essential for restoration success in post-mining areas. Previous studies have shown that soil microbial community succession of bacterial, archaeal and, fungal communities were assessed by tag encoded 454 pyrosequencing. At the phylum level, *Proteobacteria*, *Crenarchaeota*, and *Ascomycota* had the highest detected relative abundance within bacteria, archaea, and fungi, respectively (Li et al. 2016).

Soil health, and the closely related terms of soil quality and fertility, is considered one of the most important characteristics of soil ecosystems. The integrated approach to soil health assumes that soil is a living system and soil health results from the interaction between different processes and properties, with a strong effect on the activity of soil microbiota. All soils can be described using physical, chemical, and biological properties, but adaptation to environmental changes, driven by the processes of natural selection, are unique to the latter one

(Frac et al. 2018).

Management practices significant impact on the composition of the soil microbes, and changes in the microbial community composition or activity could have immediate or long-term effects on the functioning of the ecosystem (Gomez et al. 2016). Changes in the microbial community can precede detectable changes in soil physical and chemical properties, thereby providing early signs of environmental stress or ecological environment evolution in the post-coal mining sites (Li et al. 2014). The presence of soil microbes can increase the ability to return to the natural conditions of a degraded land ecosystem due to coal mining activities (Birch et al. 1991). It is important to study microbial diversity not only for basic scientific research but also to understand the link between diversity and community structure and function (Meliani et al. 2012). Fungal rhizosphere regulates important roles in ecosystem processes, however, the response of soil fungal communities to post-coal mining reclamation has not been well understood. The objective of this study was to determine the microfungus rhizosphere and soil chemical properties in a post-coal mining reclamation soil during the initial stage of plant community development.

MATERIALS AND METHODS

Study area

The materials used in this study were soils from the Embalut Coal Mining Site in Embalut Village, Tenggarong Seberang Sub-District, Kutai Kartanegara District, East Kalimantan Province, Indonesia (Figure 1). Soil samples were collected from five reclaimed areas and no

disturbance as the control. The minimum and maximum daily temperatures were 27° to 35°C, the mean monthly humidity was 91% and the mean monthly precipitation was 176.2 mm. Laboratory analysis was conducted in the Soil Science Laboratory of Tropical Rain Forest Research Center and Plant Protection Laboratory, Faculty of Agriculture, Mulawarman University. A sampling method was used to take the samples in a research location which represented the condition of every location. The reclaimed coal mining site varied in age, soil characteristics, vegetation, and type of post-mining revegetation treatment (Figure 2). A summary of each reclaimed coal mining sites is presented in Table 1.

Procedures

The samples of soil are through six different stages of reclamation, namely: undisturbed, two years after reclamation, nine-years after reclamation, twelve years after reclamation and sixteen years after reclamation (Table 1). For each stage of reclamation, three plots of 10 m x 10 m were made. Once coordinate points were determined using GPS, numbers of fungal propagules and soil physicochemical properties were measured.

Soil sampling

Rhizosphere soil samples were collected from the plots with ground drilling at a depth of 0-30 cm. On each plot, five-point drilling was determined and soil samples were composited. 500 g composite soil samples were taken from each class of depth, put into plastic bags, labeled and transported to the laboratory. Fungal propagules were determined from fresh soil samples. Soil physicochemical properties were determined from air-dried soil samples.

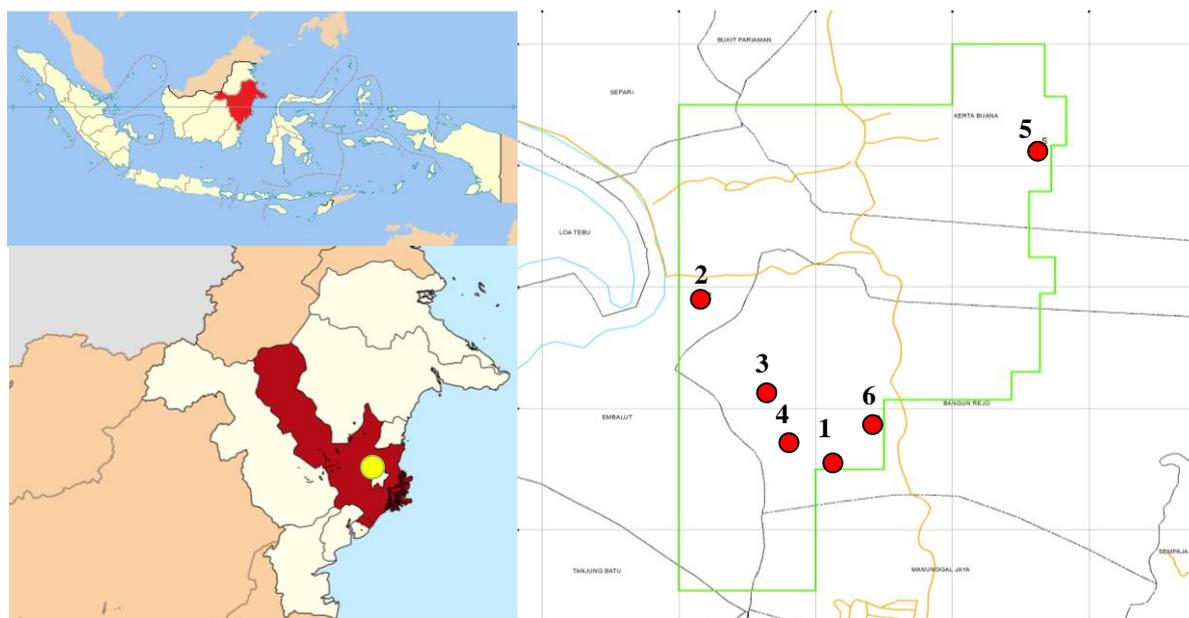


Figure 1. Location of the Study area in Embalut Village, Tenggarong Seberang Sub-District, Kutai Kartanegara District, East Kalimantan Province, Indonesia (red circles are reclamation areas). 1. Undisturbed area, 2. Disturbed area, 3. Two years after reclamation, 4. Nine years after reclamation, 5. 12 years after reclamation; 6. 16 years after reclamation.

Table 1. Description of the study sites in the Embalut Coal Mining Site, Kutai Kartanegara District, East Kalimantan, Indonesia

Plot (age in years)	Coordinate	Vegetation
Undisturbed	S 0°21'29.4444" E 117°6'7.2864"	<i>Stenotaphrum secundatum</i> , <i>Ageratum conyzoides</i> , <i>Asystasia gangetica</i> , <i>Chromolaena odorata</i>
Disturbed	S 0°20'8.484" E 117°5'10.3632"	<i>Paspalum conjugatum</i> , <i>Cynodon dactylon</i> , <i>Cyperus odoratus</i> , <i>Eclipta prostate</i> , <i>Chromolaena odorata</i>
Two years after reclamation	S 0°20'55.5144" E 117°5'39.0624"	<i>Senna siamea</i> , <i>Duabanga moluccana</i> , <i>Terminalia catappa</i> , <i>Morinda</i> <i>citrifolia</i> , <i>Samanea saman</i>
Nine years after reclamation	S 0°21'18.288" E 117°5'47.1696"	<i>Senna siamea</i> , <i>Samanea saman</i> , <i>Enterolobium cyclocarpum</i> , <i>Hibiscus</i> <i>tiliaceus</i> , <i>Gliricidia sepium</i>
Twelve years after reclamation	S 0°18'52.092" E 117°7'37.4664"	<i>Enterolobium cyclocarpum</i> , <i>Samanea saman</i> , <i>Acacia mangium</i> , <i>Gmelina arborea</i>
Sixteen years after reclamation	S 0°21'9.5436" E 117°6'33.7608"	<i>Senna siamea</i> , <i>Enterolobium cyclocarpum</i> , <i>Gliricidia sepium</i>

**Figure 2.** Illustration of sampling areas in the Embalut Coal Mining Site, Kutai Kartanegara District, East Kalimantan Province, Indonesia. A. Undisturbed area; B. Disturbed area; C. Two years after reclamation; D. Nine years after reclamation; E. 12 years after reclamation; F. 16 years after reclamation

Fungal rhizosphere counting

Fungal rhizosphere propagules were estimated using serial dilution method and plated on potato dextrose agar. Soil samples (1 g) were added to 10 ml sterilized distilled water and shaken for 15 min. The original suspension (1 ml) was diluted in 9 ml of sterilized distilled water. Series of soil dilutions of 1:10, 1:100, 1:1,000 were prepared. Each soil suspension was spread in triplicate series. Plates were incubated at 240°C under light/dark cycles for 3–4 days before measurement. Data from triplicate readings were expressed as colony forming unit (CFU) g⁻¹ of dry soil. After the measurement, isolates of the original specimens were transferred to potato dextrose agar with chloramphenicol (300 mg l⁻¹) slant tubes. The fungal isolates were identified to the genus level and in some

cases, species level, using standard keys based on morphological features (Barnett and Hunter 1972; Domsch et al. 1980). Percentage of the relative abundance of the fungal species was calculated using the following formula:

$$\% \text{ of relative abundance} = \frac{\text{Total number of colonies of a particular genus/species}}{\text{Total number of colonies of all the genera / species}} \times 100$$

Soil analysis

The soil sample was taken from the depth of 0-30 cm and the results were calculated after being dried in the oven with the temperature of 150°C until constant weight was reached. Three major soil nutrient components and other two soil properties, namely soil pH, organic carbon, total

nitrogen, available phosphorus, and available potassium, were measured. The pH was determined using a pH meter (water-to-soil ratio 1:2.5); soil organic carbon was determined by the Walkley-Black method; total N was determined by Kjeldahl determination; available P was determined using Spectronic method; and available K was determined using Atomic Absorption Spectrophotometry.

Data analysis

Variations in fungal rhizosphere propagules and the soil physicochemical properties were analyzed using standard analysis of variance. Statistical analysis was undertaken using SPSS statistical software version 21.0. All data were expressed as the mean \pm standard deviation. The Least Significant Difference (LSD) test was used to compare fungal propagules and soil physicochemical properties mean at the 5% probability level.

RESULTS AND DISCUSSION

Soil characteristics

Table 1 shows that there was a significant variation ($p < 0.05$) in the chemical properties among the reclamation stages of different age groups. The pH values of samples ranged from 6.27 to 8.47 indicating that they were neutral and moderately alkaline. The concentrations of organic carbon status were medium (2.57 to 2.81%) to high (3.69 to 3.75%). Organic carbon concentrations in the 9 and 12 years after reclamation were significantly higher than the undisturbed soil and the older reclamation soil. The concentration of organic carbon increased in 2 years after reclamation and then decreased after 9 years after reclamation. Total N ranged from 0.08 to 0.18% including low and very low categories. This study shows that two years after reclamation soil had significantly lower total N concentration than the undisturbed soil of the reclaimed sites. The concentration of available P has the status of extremely low (1.69 to 4.18 ppm) to medium (6.37 to 8.26 ppm). The available P concentrations in the nine years after reclamation soil were significantly higher than the disturbed soil and the youngest reclamation soil. The concentration of available K status was low (74.04 to

155.69 ppm). Considering all the reclamation stages, the twelve years after reclamation stage exhibited significantly lower available K concentration indices than other reclamation stages. The concentration of total N, available P and available K generally increased in the early years after reclamation but decreased slightly in more than ten years after reclamation. The contents of Ca, Mg, Fe, Mn and S for disturbed area was higher than the undisturbed area indicating that coal mining and soil process reclaimed may affect the metal exposed from the soil.

Composition and diversity of understory vegetation

The results of the observation show that, among the study sites, there were differences total plant species and in importance value of index species composition and familial families in the land stands after coal mining reclamation (Fig 3 and Table 2). From Table 2, the amount of IVI is known and there is a predominance of three understory species in the study sites, namely *Paspalum conjugatum*, *Asystasia gangetica*, and *Stenotaphrum secundatum*. *Paspalum conjugatum* was dominant in active mining sites in 2 and 16 years after coal mining reclamation. *Asystasia gangetica* was dominant inland in 9 and 12 years after coal mining reclamation. While the type of understory in non-mining land was dominated by the *Stenotaphrum secundatum*. The dominance of the three species of understory was because they belong to a group of clumps that are able to breed in large quantities and quickly dominate a land community compared to other plant species.

The results of the Shannon-Wiener (H') diversity index analysis from each study site can be seen in Figure 4. Land that has been reclaimed showed a relatively higher diversity index value compared to non-mining land and active mining. The results of the Pielou (e) evenness index analysis formed a pattern similar to the Shannon-Wiener (H') diversity index that has been reclaimed shows a relatively higher Pielou evenness index compared to non-mining land and active mining. The Pielou (e) evenness index value in non-mining land is relatively low compared to other locations due to the dominance of certain species despite relatively high species and family wealth.

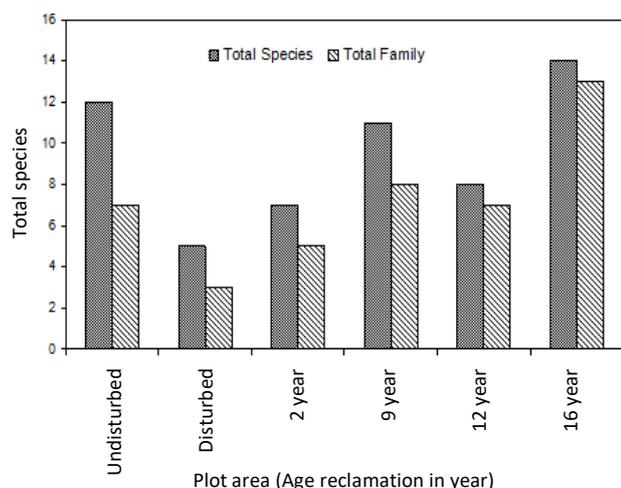
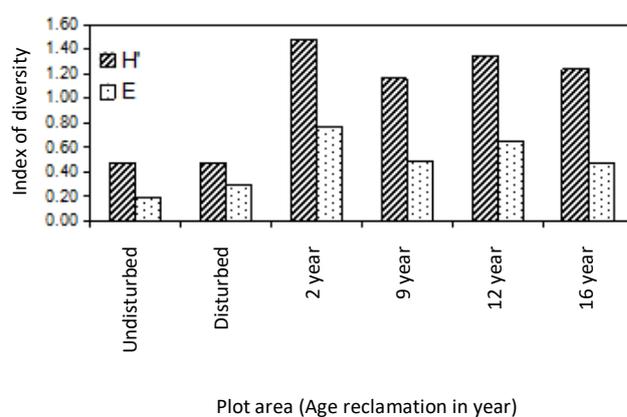
Table 1. Physicochemical properties of different reclaimed ages

Parameter	Plots (years after reclamation)					
	Undisturbed	Disturbed	2 year	9 year	12 year	16 year
pH	6.27 \pm 1.78 ^a	7.85 \pm 0.1 ^{bc}	8.47 \pm 0.07 ^c	6.12 \pm 0.72 ^a	6.96 \pm 0.49 ^{abc}	6.82 \pm 0.20 ^{abc}
SOM (%)	2.57 \pm 0.15 ^a	3.69 \pm 0.72 ^b	2.81 \pm 0.17 ^a	3.75 \pm 0.72 ^b	3.73 \pm 0.09 ^b	2.67 \pm 0.36 ^a
Total N (%)	0.14 \pm 0.02 ^b	0.13 \pm 0.02 ^{ab}	0.08 \pm 0.03 ^a	0.18 \pm 0.05 ^b	0.17 \pm 0.04 ^b	0.13 \pm 0.02 ^{ab}
Available P (ppm)	3.49 \pm 0.71 ^{abc}	1.69 \pm 1.4 ^a	2.81 \pm 1.11 ^a	8.26 \pm 1.1 ^d	6.37 \pm 3.65 ^{cd}	4.18 \pm 0.71 ^{abc}
Available K (ppm)	127.55 \pm 18.78 ^c	147.33 \pm 8.21 ^d	149.35 \pm 2.51 ^d	155.69 \pm 2.0 ^d	74.04 \pm 2.31 ^a	92.02 \pm 0.96 ^b
Ca (meq.100 g-1)	1.53 \pm 0.91 ^a	8.12 \pm 0.51 ^d	4.57 \pm 0.36 ^c	3.52 \pm 0.33 ^b	5.12 \pm 0.72 ^c	9.16 \pm 0.43 ^e
Mg (meq.100 g-1)	1.83 \pm 1.32 ^a	6.28 \pm 0.29 ^c	4.43 \pm 0.32 ^b	4.73 \pm 0.46 ^b	6.14 \pm 0.77 ^c	8.55 \pm 0.19 ^d
Fe (mg.kg ⁻¹)	2,404.04 \pm 47.57 ^a	2,380.95 \pm 21.7 ^a	2,315.04 \pm 17.10 ^a	2,280.11 \pm 17.42 ^a	2,356.72 \pm 23.74 ^a	2,380.38 \pm 23.92 ^a
Mn (mg.kg ⁻¹)	312.31 \pm 123.45 ^{ab}	754.78 \pm 20.10 ^d	474.51 \pm 15.54 ^{bc}	175.81 \pm 77.05 ^a	494.55 \pm 192.69 ^{bc}	666.40 \pm 90.38 ^{cd}
S (%)	4.91 \pm 0.01 ^a	39.76 \pm 15.89 ^c	13.39 \pm 4.10 ^{ab}	29.15 \pm 13.49 ^{bc}	27.63 \pm 13.39 ^{bc}	13.09 \pm 0.019 ^{ab}

Note: n=3, mean SE) of soil for each site. Values followed by different letters (a, b, c, d, e) between location are significantly different ($P < 0.05$) according to Duncan's multiple comparison. SOM, Soil organic matter; Total N, Total nitrogen; Available P; Available K, Available potassium

Table 2. The results of importance value index of understorey vegetation on land post-coal mining reclamation

Species	Family	Importance Value Index (IVI)					
		Undisturbed	Disturbed	2 year	9 year	12 year	16 year
<i>Ageratum conyzoides</i>	Asteraceae	13.71			7.7	20.27	6.74
<i>Chromolaena odorata</i>	Asteraceae	12.20	11.5				
<i>Mikania micrantha</i>	Asteraceae	5.38			6.63		
<i>Eclipta prostrata</i>	Asteraceae	24.88					
<i>Chromolaena odorata</i>	Asteraceae			13.84			
<i>Eupatorium odoratum</i>	Asteraceae						11.51
<i>Melastoma malabatricum</i>	Melastomaceae	11.14			6.0		
<i>Paspalum distichum</i>	Poaceae	5.71	122.71	65.34			84.54
<i>Pennisetum purpureum</i>	Poaceae	5.43			87.10		
<i>Isachne globosa</i>	Poaceae	5.43					
<i>Stenotaphrum secundatum</i>	Poaceae	106.72					
<i>Cynodon dactylon</i>	Poaceae	26.65					
<i>Paspalum distichum</i>	Poaceae					11.60	
<i>Ischaemum timorense</i>	Poaceae			23.37		19.4	
<i>Digitaria sanguinalis</i>	Poaceae				26.8		
<i>Fimbristylis miliacea</i>	Cyperaceae	5.49					
<i>Cyperus odoratus</i>	Cyperaceae		14.21	11.46			
<i>Cyperus rotundus</i>	Cyperaceae				6.63		
<i>Asystasia gangetica</i>	Acanthaceae	17.47				79.68	25.33
<i>Calopogonium mucunoides</i>	Fabaceae	5.88					5.05
<i>Pueraria javanica</i>	Fabaceae			60.58			7.31
<i>Mimosa pudica</i>	Fabaceae			13.84			
<i>Solanum torvum</i>	Solanaceae			11.46	21.5		11.22
<i>Capsicum frutescens</i>	Solanaceae				6.0		
<i>Nephrolepis falcate</i>	Nephrolepidaceae				6.09	32.96	10.94
<i>Matteuccia struthiopteris</i>	Onolaceae				10.93		
<i>Piper aduncum</i>	Piperaceae					16.37	5.33
<i>Taenitis blechnoides</i>	Pteridaceae					11.60	5.90
<i>Urena lobata</i>	Malvaceae						5.05
<i>Passiflora foetida</i>	Passifloraceae						7.31
<i>Baccaurea polyneura</i>	Phyllanthaceae					8.19	
<i>Phyllanthus niruri</i>	Phyllanthaceae						6.74
<i>Cyclosorus interruptus</i>	Thelypteridaceae						7.03

**Figure 3.** Total plant species at different age post-coal mining reclamation sites**Figure 4.** Shanon-Wiener Diversity index (H') and Pielou evenness index (E) of understorey vegetation on Post-reclamation Area

Fungal rhizosphere communities

Numbers of fungal propagules were significantly different in the undisturbed soil, the disturbed soil or the reclaimed soil ($P < 0.05$) (Table 3). Numbers of fungal propagules in the undisturbed soil differ with the post-coal mining reclamation. Fungal propagules show increased in the year after reclamation but not significant and then decreased after twelve years of reclamation compared to undisturbed soil. This result indicates that fungal rhizosphere propagules were different in soils under different vegetation covers. The decline in fungal propagules probably reflected the exhaustion of added carbon and nitrogen from the litter. Each site had a different fungal propagule, thus indicating that a given fungal group had a close relationship to a specific plant successional stage of vegetation.

Distribution number of fungal genera in the various stages of reclamation can be seen in Table 4. The result shows that many fungal genera isolated from the reclamation of coal mining sites were common to all areas. In this study, the important fungal genera with respect to the number of propagules isolated were *Aspergillus*, *Penicillium*, *Cephalosporium*, *Gliocladium*, *Humicola*, *Trichoderma*, and *Paecilomyces* (Figure 5). They were among the fungi found colonizing in the soil and the most diverse and ecologically significant group of fungi on the rhizosphere. However, some appeared in considerably higher densities than others. *Aspergillus* was the dominant genus that comprised 74% of the total isolates from these reclamation areas. After nine years of reclamation, the soil had higher fungal propagules but lower fungal diversities than disturbed soil.

Relationship between plant diversity and abundance of soil fungi

The results of correlation test show that the Shannon-Wiener diversity index (H') and Pielou (E) evenness of plants have a weak positive correlation with the abundance of soil fungi (Figure 6 and Figure 7). A low correlation coefficient is caused by very diverse environmental conditions. Thus, the abundance of soil fungi on post-coal mining reclamation soil is not only related to plant diversity but also related to environmental conditions.

The results of this study have not been able to show the factors related to the abundance of soil fungal populations, especially their association with the existence of plants on post-coal mining reclamation; however, plant analysis and soil sampling have been carried out specifically on the same plot. The existence of plants in the location of observation is suspected to be less diverse. Therefore, further research with a higher level of diversity of plants is still needed to identify the plant diversity with abundance of land after coal mining reclamation. This is because plants have a role as a bridge between the ecosystem above and in the soil. Turnbull et al. (2016) stated that ecosystems function better when they contain more species. Garcia-Palacios et al. (2015) state that changes in plant structure will affect soil conditions, including processes of soil formation, soil structure and microbial community of soil,

in this case, the abundance of soil fungi. This is supported by Widyati (2013) who stated that plants influence the soil microbial community through the provision of C through root exudates. As a result, the activity and number of microorganisms around the roots (rhizosphere) will be far greater than the surrounding soil. Different plants will produce different types and compositions of exudates that will determine the composition of the diversity of soil microbial communities.

Discussion

The results presented in Figure 2 show that the fungal rhizosphere propagules in the undisturbed soil and the post-coal mining reclamation soil were higher compared to the disturbed soil. The changes in the ecosystem structure have an impact on the composition of the fungal propagules in the reclamation areas. Our interpretation is that the dynamics of the fungal propagules observed seem to be related to changes in the availability of carbon resources occurring during degradation, and that the low numbers of fungal propagules could have been due to the absence of organic matter in the form of litter (Frezquez et al. 1987; Gomez et al. 2016).

Table 3. Density of fungal rhizosphere propagules Σ colony (CFU.g⁻¹)*10³ in various stages of reclamation

Years after reclamation	Density of soil fungi Σ colony (CFU.g ⁻¹)*10 ³			Mean
	I	II	III	
Undisturbed	130	75	105	103.33 ±15.89 ^{ab}
Disturbed	10	85	15	36.67 ±24.20 ^a
2-year	125	40	90	85.00 ±24.20 ^{ab}
9-year	110	170	80	120.00 ±26.45 ^b
12-year	75	40	120	78.33 ±23.15 ^{ab}
16-year	130	84	45	86.33 ±24.56 ^{ab}

Note: Values followed by different letters(a,b,c,d,e) between location are significantly different ($P < 0.05$) according to Duncan's multiple comparisons

Table 4. Distribution number of fungal genera in the various stages of reclamation

Fungal groups	Un-disturbed	Disturbed	Years after reclamation			
			2-year	9-year	12-year	16-year
<i>Aspergillus</i>	7	6	5	6	4	6
<i>Penicillium</i>	1	0	1	1	0	1
<i>Cephalosporium</i>	0	1	0	0	0	0
<i>Gliocladium</i>	0	1	0	0	1	0
<i>Humicola</i>	0	0	0	0	1	0
<i>Trichoderma</i>	0	0	0	0	1	0
<i>Paecilomyces</i>	0	0	1	0	0	0
Unidentified						
Isolate No. 1	1	0	0	0	0	0
Isolate No. 2	0	0	0	0	1	0
No. of isolates	9	8	7	7	8	7
No. of genera	3	3	3	2	5	2

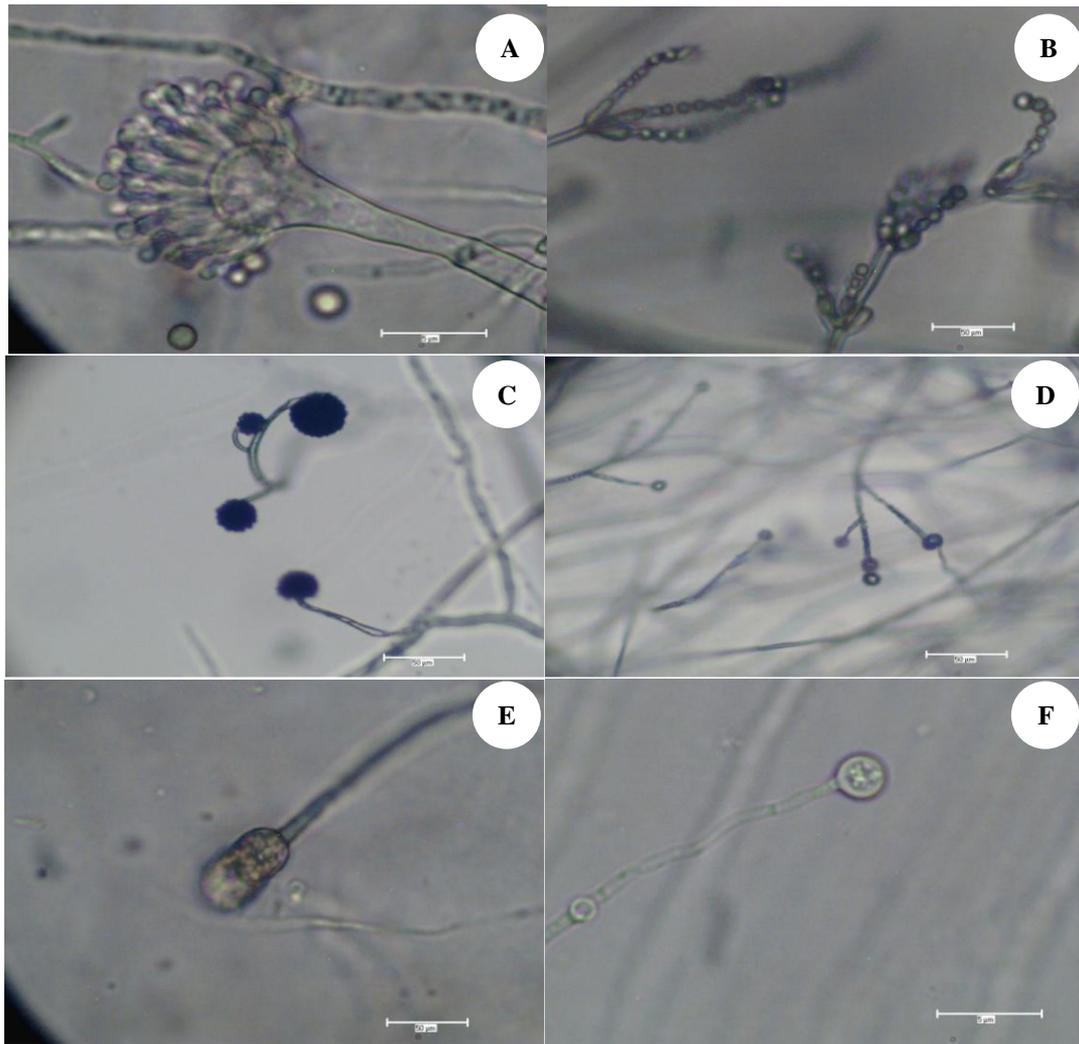


Figure 5. Several types of soil fungi identified from post-coal mining reclamation soil. A. *Aspergillus* sp.; B. *Penicillium* sp.; C. *Gliocladium* sp.; D. *Trichoderma* sp.; E. Not yet identified; F. Not yet identified

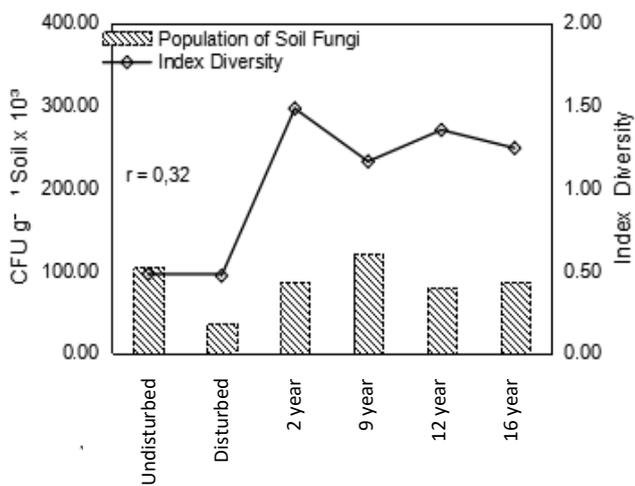


Figure 6. Relationship between index of plants diversity with abundance of soil fungi on post-reclamation

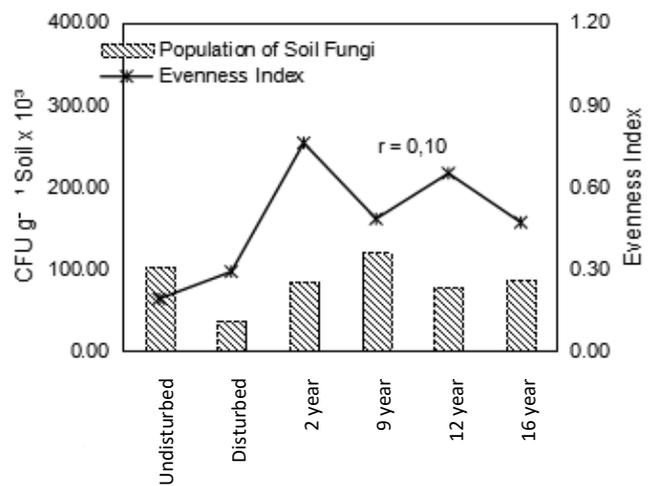


Figure 7. Relationship between index of plants diversity with abundance of soil fungi on post-coal mining reclamation

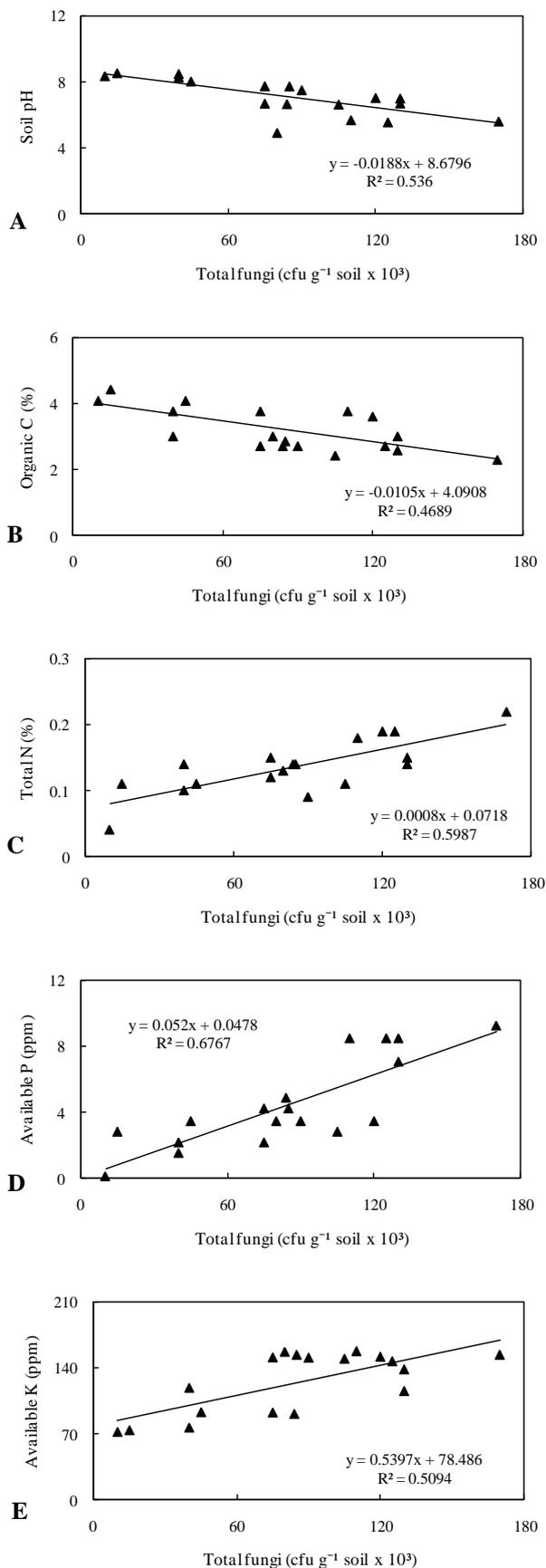


Figure 8. Relationship between fungal rhizosphere with: A. Soil pH, B. Organic carbon, C. Total N, D. Available P, E. Available K at reclamation areas

We also expected that changes in fungal propagules would be strongly linked to vegetation dynamics. Our results show that the numbers of fungi propagules differed appreciably from one ecological area to the others which were influenced by vegetation cover and especially by soil factors. Our data indicate that fungal rhizosphere propagules were different in soils under different vegetation covers. Roy-Bolduc et al. (2016) found a significant correlation between vegetation and fungal rhizosphere communities which were influenced by similar soil physicochemical properties. According to Setiawati (2014), the numbers of soil microbes found at plant rhizosphere were related to root exudate. Some researches showed that root exudate plays a central role in influencing interactions with neighboring plants and microbes. The quantity and quality of root exudate depend on plant species, developmental stage, and environmental factors. Garcia et al. (2015) reported a similar result. Meliani et al. (2012) stated that the activity and species composition of microbes were generally influenced by many factors including soil physicochemical properties, temperature, and vegetation.

The genera with a higher dominance in the total fungi isolated, according to the number of the genus, were *Aspergillus* (74%), *Penicillium* (8%), *Gliocladium* (4%), *Cephalosporium*, *Gliocladium*, *Humicola*, *Trichoderma*, *Paecilomyces*, and unidentified genera (2%). The results of this study showed that the reclamation areas were dominated by *Aspergillus* and *Penicillium* (Table 2). Kurakov et al. (2008) reported similar fungal genus in their work, and Garcia et al. (2015) stated that these genera are capable of using different substrates in the soil, they predominated over other fungal propagules that used only a few specific substrates. Domsch et al. (1980) stated that *Aspergillus* and *Penicillium* can utilize a wide range of substrate and can survive under extreme environmental conditions. Gomez et al. (2016) stated the reason why other fungal genera were in a low proportion or absent in the sampling areas was that saprophytic fungi present in a high relative density. After nine years of reclamation, the soil had higher fungal propagules showed lower fungal diversities than disturbed soil. The higher fungal propagules and lower fungal diversities are related to either adverse soils or surface conditions, or to soils high in nutrients levels. Frezquez et al. (1987) described that the composition of fungal rhizosphere is related to the properties of the soil, as well as to the type of vegetative cover during succession. Generally, a high diversity of fungal species and a few individual organisms per species are indication of stable and well-balanced soil ecosystems. On the contrary, low fungal diversity with high numbers of individual organisms per species indicate unstable or stressed soil ecosystems.

The results of this study indicated that six stages of reclamation significantly altered the physicochemical properties of post-coal mining sites (Figure 8). The soil pH after two years of reclamation was significantly higher than the undisturbed soil in any of the reclamation areas. Organic carbon concentrations after nine years of reclamation were significantly higher than the undisturbed

soil and the later reclamation stages. The concentration of total N, available P, and available K generally increased as the year went up to the nine years after reclamation but decreased slightly in the later reclamation stages. The accumulation of many soil physicochemical properties was found to be higher in the nine years after reclamation compared to any of the reclamation areas. As described by Frezquez et al. (1987), these results suggest that the reclamation areas may not yet be stable since there may still adverse soil chemical and physical properties (i.e. high salts and high clay contents). Studies have shown significant interactions between soil and vegetation during a reclamation process, such that soil and vegetation are constantly evolving and developing (Frezquez et al., 1988; Meliani et al. 2012; Yang et al. 2016). Rousk et al. (2009) describe fungi and bacteria are the two groups that dominate the microbial decomposer community, and, crudely define that they share the function of decomposing organic matter in the soil, indicating that there is a strong potential for interaction. More recently, Garcia-Palacios et al. (2015) and Ashworth et al. (2017) reported that soil microbial communities are vital for continuous cycling of nutrient and for driving aboveground ecosystems. Vegetation cover and fungal rhizosphere propagules varied among six reclamation stages, leading to the differences in both mineralization of soil organic matter and soil physicochemical properties.

The fungal rhizosphere propagules correlated negatively with soil pH and organic carbon. The slightly higher fungal propagules in the reclamation areas can be attributed to lower soil pH and organic carbon concentrations. The relation of fungal propagules to soil pH demonstrated that fungal propagules increased as pH decreased. These results are in agreement with the previous reports by Rousk (2009) and Udom and Benwari (2018), focusing on the effects of fungal growth and biomass on soil pH. In this study, soil chemical analysis shows that the soil pH ranged between 6.27 to 8.47, generally, fungal growth was optimal at pH 4.5 to 5, and then decreased above pH 5 (Suciatmih 2006; Setiawati 2014). Our results also indicate that higher fungal propagules can be attributed to lower organic carbon concentrations. Hrselova et al. (1999) reported the similar results in their work, the concentration of CFU of culturable fungi were negatively correlated with organic carbon observed in field-collected soil samples.

On the other hand, the increase of fungal propagules of the soil was positively correlated to the increase of total N, available P, and available K indicating that total N, available P, and available K increased as the fungal propagules with a range of about 3.7×10^4 to 1.2×10^5 CFU g^{-1} soil. As described by White et al. (1997), fungi can withstand a much wider pH range, and many can produce organic acids which can solubilize and boosts the availability of N, P, K and Fe in the soil. A lot of fungi are capable of producing organic acids, including *Aspergillus niger* (citric, oxalic, gluconic), *Aspergillus* spp. (citric, malic, tartaric, α -ketoglutaric, itaconic, aconitic), and *Penicillium* spp. (citric, tartaric, α -ketoglutaric, malic, gluconic). By altering parameters such as the N, P and K

balance of the soil and optimum pH, these species can be used to produce a wide range of organic acid for the mobilization of nutrients and facilitate their plant uptake from the rhizosphere.

The presence of fungal rhizosphere is an important agent to improve the availability of total nitrogen. As discussed above, fungi indirectly affect N fixation through bacteria present in mycelia. During this process, fungi translocate carbon and P from the plant roots to the associated bacteria for N fixation (Rashid et al. 2016). Setiawati (2014) stated that some fungal rhizospheres are capable to dissolve soil P continuously. Their activity increases P availability through several mechanisms, i.e., mineralization of organic P by phosphatase enzyme to an organic P, which can be used by the plants, competition of adsorption site, and changes of soil reaction. Rashid et al. (2016) described the major processes involved in mobilizing K are acidolysis and complex lysis exchange reactions. During acidolysis, soil microbe such as fungi decreases the local pH by producing succinic, citric, gluconic, α -keto-gluconic and oxalic acids. This process plays an important role in mobilizing or solubilizing insoluble form and structural unavailable forms of K compounds into soil solution resulting in an increased K availability in the rhizosphere.

In conclusion, the present study results show that nine isolates of microfungi were identified from the six stages of reclamation samples in post-coal mining reclamation sites, i.e., *Aspergillus*, *Penicillium*, *Cephalosporium*, *Gliocladium*, *Humicola*, *Trichoderma*, *Paecilomyces* while two other genera are unidentified yet. The genera with a higher dominance in the total fungi isolated, according to the number of the genus, were *Aspergillus* (74%), *Penicillium* (8%), *Gliocladium* (4%), *Cephalosporium*, *Gliocladium*, *Humicola*, *Trichoderma*, *Paecilomyces*, and unidentified genera (2%). Numbers of fungal propagules were not significantly different in the undisturbed soil, the disturbed soil or the reclaimed soil. Fungal propagules increased in the early year after reclamation and then decreased after twelve years of reclamation comparable to those of the undisturbed soil. The fungal rhizosphere propagules were different in soils under different vegetation covers. Fungal populations and community composition were related to soil properties and site age of reclamation. The fungal propagules were negatively correlated with soil pH and organic carbon. On the contrary, fungal propagules to be positively correlated with total N, available P, and available K.

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