Abstract. Budi SW, Wibowo C, Sukendro A, Bekti HS. 2020. Growth improvement of Falcataria moluccana inoculated with MycoSilvi grown in post-mining silica sand soil medium amended with soil ameliorants. Biodiversitas 21: 422-427. High aluminum content in soil of post-mining silica sand area inhibits plant growth. MycoSilvi is an inoculum of Arbuscular Mycorrhizal Fungi (FMA) enriched with Mycorrhizal Helper Bacteria (MHBs) which plays an important role for improving plant growth in unfertile soil medium. The aims of this research were to analyze the growth response of Falcataria moluccana (Miq.) Barney & JW Grimes) seedlings treated with MycoSilvi and soil ameliorants (compost and lime) in post-mining silica sand soil medium. The randomized complete design with factorial scheme was used in this study. The results showed that the interactions of MycoSilvi and Soil ameliorant significantly increased height, diameter, and mycorrhizal colonization of F. moluccana. Combination of MycoSilvi variant 3 and lime increased height, diameter, and biomass of F. moluccana by 96.5%, 147%, and 1427% respectively, as compared to those of control plants. The mycorrhizal roots colonization in those treatments was 98%. The addition of compost and lime increased pH and decreased Aluminum and Fe of the soil medium. F. moluccana seedlings have high mycorrhizal dependency on post-mining silica sand soil media. These results indicate prospective uses of MycoSilvi and soil ameliorants for improving plant growth in unfertile soil medium, including soil in post-mining area.

Keywords: Aluminum, Falcataria moluccana, MycoSilvi, post mining, soil ameliorant

INTRODUCTION

Mining activities are mostly carried out by open mining method, which causes scraping of the upper soil surface. This open mining system will cause deterioration of soil properties, including physical properties, chemical properties and biological properties of the soil (Arshi 2015). Changes in soil properties can result in land degradation as indicated by structural indicators such as low total plant cover, low perennial and annual plant species richness, low aboveground Phytomass, low beta diversity, decreased life form spectrum, reduced number of keystone species, slow microbial biomass, low soil microbial diversity, as well as functional indicators such as low biomass productivity, low soil organic matter and poor soil water retention (Asmelash et al. 2016). Degraded lands are also characterized by low levels of Arbuscular Mycorrhizal Fungi (AMF) abundance and diversity (Cardozo-Junior et al. 2012). Plants can not grow normally in degraded land as reported by Widyati (2006) due to low pH and solubility of heavy metals.

The solubility of heavy metal, especially Al and Fe in the soil, greatly correlated with soil pH (Delhaize and Ryan 1995). When soil pH was bellow 5.5, Al³⁺ become dominant and constituted the most toxic form for plant growth and development (Kochian et al. 2004). Al³⁺ element is a limiting factor in plant growth and development, and caused phenomena such as stunted root growth and root extension to take soil nutrients (Silva 2012). Toxic elements cause deviations of physiological and biochemical processes during plant growth (Barchia 2009). The presence of high Al and Fe content in post-mining land have been reported by Jayani et al. (2018) and inhibited the early growth of three species of forest seedlings.

Several soil ameliorants have been used by many researchers for improving plant growth and development in acidic soil medium (Ohsowski et al. 2017; Mrabet et al. 2014; Moreira et al. 2010). The use of lime for increasing soil pH has been reported by several researchers (Mrabet et al. 2014, Jayani et al. 2018; Guo and Huang 2010). The use of compost and biochar for improving plant growth and development in post-mine sandpits were well documented (Ohsowski et al. 2017). Some researchers used Arbuscular Mycorrhizal Fungi for improving plant growth in acidic soil medium (Husna et al. 2015; Husna et al. 2019) and in heavy metal contaminated soil (Setyaningsih et al. 2018). In the rhizosphere, AMF interacts positively with biochar and give synergetic effect to plant growth (Budi and Christina 2013; Budi and Setyaningsih 2013). Positive interaction of compost, lime, and AMF which was formulated as MycoSilvi inoculum, containing Glomus mosseae and Mycorrhizal Helper Bacteria (MHBs) have been reported by Jayani et al. (2018) and can improve early growth of forest trees seedling grown in post-mine soil medium.

In this research, we develop a new variant of MycoSilvi which consists of three variants, namely MycoSilvi variant
1 containing Glomus mosseae, MycoSilvi variant 2 containing G. mosseae and Acaulopora sp, and MycoSilvi variant 3 containing G. mosseae, Acaulopora sp., and Gigaspora margarita. We hypothesize that MycoSilvi containing more than one AMF species will be more effective than one AMF species. The aim of this study was to analyze growth response of Falcataria moluccana seedlings inoculated with different variants of MycoSilvi, and the seedlings were grown in post silica sand-mine soil medium amended with lime and compost.

MATERIALS AND METHODS

Germination of Falcataria moluccana seeds

The experiment was carried out in greenhouse at the Department of Silviculture, Faculty of Forestry, IPB University, Bogor, Indonesia. F. moluccana seeds, obtained from the Center of Research and Development for Forest Seed, Bogor Indonesia, were soaked in hot water (80°C) for one hour and were further soaked in cold water for 24 hours for breaking their dormancy. The seeds were then sown in a plastic box, containing sterile zeolite and watered daily as needed.

Preparation of soil growth medium

The soil medium was collected from silica post-mining area of PT Holcim Indonesia, Tbk, Sukabumi, West - Java. The soil medium was then air-dried, sieved with soil sieve of 2 mm and autoclaved at 1.5 psi, 120°C for one hour. The physicochemical characteristics of soil medium have been analyzed at Soil Laboratory, Faculty of Agriculture, IPB University comprising the following variables: pH (H₂O) (3.56); C-organic (1.65%); N-total (0.29%); P-total (46.18 mg/100g); P-available (4.25 mg/100g); Ca (0.26 cmol/kg); Mg (0.29 cmol/kg); K (0.10 cmol/kg); Na (0.04 cmol/kg); CEC (11.02 cmol/kg); Al (5.75 cmol/kg); Fe (182.98 ppm). The soil showed the following soil texture: Sand 37.95%; Silt 27.01% and Clay 35.04%. Lime was obtained from farmer market as dolomite. Organic compost was obtained from CV. Cahaya Gemilang, Bogor Indonesia and have the following chemical characteristics: pH (H₂O) (7.61); C-organic (47.14%); N-total (1.10%); ratio C/N (42.68); P₂O₅ (1.67%); K₂O (2.44%) and moisture content (15.66%). Sterilized soil medium was then mixed or not with organic compost and lime, and filled to polybag 15 x 20 cm.

Preparation of MycoSilvi inoculum and inoculation of plants

MycoSilvi inoculum was produced in plastic pot of 1000 ml containing zeolite sterile medium by using Pueraria javanica as host plants for two months. At harvest, the number of AMF spores per gram medium was counted and stored in refrigerator until used.

_F. moluccana_ seedlings that had aged two weeks in plastic box containing zeolite sterile medium were transferred in to polybag filled with sterilized growing medium. MycoSilvi were inoculated to the healthy and uniform _F. moluccana_ seedlings. Each plant received 5 g of MycoSilvi inoculum (containing 50 spores of Arbuscula Mycorrhizal Fungi). AMF inoculation was conducted near the root system of the plants and the height and stem diameter of the seedlings was determined at the AMF inoculation as an early baseline data.

Parameters evaluation

The evaluation of variables was carried out at 12 weeks after inoculation of _F. moluccana_ seedlings, comprising the following variables: plant height, stem diameter, dry biomass, and mycorrhizal root colonization. Plant heights were determined by measuring tape, stem diameter by using digital caliper, and plant biomass was determined after drying in oven at 70°C for 72 hours. The estimation of percentage of mycorrhizal root colonization was carried out after bleaching and staining, according to the method described by Philip and Hayman (1970). The quantification of mycorrhizal roots colonization was performed by a method of Brundrett et al. (1996). The growth response (GR) of _F. moluccana_ was calculated according to the formula developed by Hetrick et al. (1996), while the mycorrhizal dependence (MD) was calculated according to the method of Plenchette et al. (1983).

Experimental design and data analysis

The experimental design was randomized complete design, in 4 x 2 x 2 factorial scheme, with 4 MycoSilvi treatments: control (not inoculated), inoculated with MycoSilvi variant 1, inoculated with MycoSilvi variant 2, and inoculated with MycoSilvi variant 3; two organic compost doses (0 g and 32.5 g)/650 g growth medium; and two lime doses (0 g and 7.2 g)/650 g growth medium. Each treatment was replicated five times. Analysis of variance was carried out, followed with Duncan test to compare all treatments.

RESULTS AND DISCUSSION

Effect of MycoSilvi and soil ameliorants on plant growth

It is well known that inhibition of plant growth is mainly due to inadequate levels of any nutrients (Lambers et al. 1998) as well as the availability of toxic elements in soil media such as Al and Fe (Silva 2012). Bioavailability and toxicity of Al occurs mainly on acid soil with pH below 5.5, and consequently decreased crop production (Alori and Fawole 2012; Silva 2012). According to Yamamoto (2019), the plant cell deaths were triggered by Aluminum due to enhancement of Fe-mediated lipid peroxidation, leading to a loss of plasma membrane integrity, dysfunction of mitochondria followed by Reactive Oxygen Species (ROS) production and vacuolar collapse due to upregulation of a vacuolar processing enzyme. In this research, the medium growth being used had low pH (H₂O) (3.56); low C-organic content (1.65%); low N-total (0.29%); low P-available (4.25 mg/100g); high Al (5.75 cmol/kg); and high Fe (182.98 ppm). Consequently, control plants without soil amendment and not inoculated with MycoSilvi showed stunted growth as compared to other treatments (Table 1). On the contrary, 12 weeks old _F. moluccana_ seedlings inoculated with
MycoSilvi significantly showed increased height, stem diameter and biomass for all MycoSilvi variants (Table 1). The plant height, stem diameter, and biomass increased differently in line with MycoSilvi variants being used, regardless of soil amendment application (Table 1). The plant heights were increased by 24%, 293%, and 575% when inoculated with MycoSilvi variant 1, 2 and 3 respectively (Table 1), while the plant stem diameter with the same treatment was increased by 79%, 91%, and 102% respectively as compared to control. The plant biomass also showed similar results when plants were inoculated with MycoSilvi variants 1, 2 and 3, which were increased by 317%, 883%, and 926% respectively as compared to control plant.

Plant growth improvement by MycoSilvi variant 3 showed better results than those by MycoSilvi variant 2 and variant 1, due to different AMF species contained in the MycoSilvi. MycoSilvi variant 1 contained one AMF species Glomus mosseae, while MycoSilvi variant 2 and 3, contained two and three AMF species, namely G. mosseae and Acaulospora sp and G. mosseae, Acaulospora sp and Gigaspora margarita respectively. These results were in agreement with Chen et al. (2017), who found that Cucumber seedling growth was better when inoculated with five AMF species than four or one AMF species. AMF has an extrametrical hyphae network in the plant rhizosphere and plays important roles not only in promoting plant growth in degraded soil (Berruti et al. 2016) but also improving soil aggregation (Borie et al. 2008) and soil chemical properties (Pal and Pandey 2017). AMF also plays an important role in nutrient uptake from soil medium, especially Nitrogen (Bucking and Kafle 2015) and Phosphorus (Goussous and Mohammad 2009). In addition, AMF also produced phosphatase enzyme and have been reported by several researchers (Bini et al. 2017; Sultana and Siddique 2015; Wang et al. 2011). This enzyme plays an important role in releasing Al/Fe-P bound in acid soil and absorbed by mycorrhizal hyphae or root plant. In this study, plants inoculated with MycoSilvi showed better growth than control plant, indicating that MycoSilvi has been functioned, as shown also by mycorrhizal roots colonization (Table 1).

The roles of organic compost for improving plant growth in acid soil have been reported by several researchers (Escober and Hue 2008; Bougnom et al. 2010; Medina and Azcon 2010). This organic material have a direct effect on reduced Al toxicity, increased soil organic matter content, increased soil pH and soil biota, increased aggregate stability, soil enzymatic activities, water-soluble C and water-soluble carbohydrates, as well as nutrient availability, especially P. In this research, addition of compost to soil medium increased plant height, stem diameter and biomass by 105%, 50%, and 113% respectively as compared to control plant (Table 1). Interestingly, addition of compost to soil medium can improve AMF growth and development as indicated by increasing percentage of mycorrhizal roots colonization (Table 1), and consequently increased plant height, stem diameter, and biomass. Interaction between MycoSilvi variant 1 and compost increased plant height, stem diameter and biomass by 961%, 131%, and 1736% respectively as compared to control plant. On the other hand, the interaction between MycoSilvi variant 1 and compost increased plant height, stem diameter and biomass by 521%, 121%, and 1347% respectively, as compared to control plant. The similar results when plant inoculated by MycoSilvi variant 3 combined with compost, can increased plant height, stem diameter, and biomass by 510%, 110% and 803% respectively as compared to control plant. Our results are contrary to those of Mrabet et al. (2014), who found that mycorrhizal plants in soil media not amended with bio-compost showed higher percentages of root colonization than those of inoculated plants planted in amended soils, and therefore bio-compost had a negative effect on AMF development. This was due to different chemical characteristics of soil medium and compost used in this study especially pH and C/N ratio.

<table>
<thead>
<tr>
<th>MycoSilvi</th>
<th>Soil ameliorant</th>
<th>Compost</th>
<th>Lime</th>
<th>Height (cm)*</th>
<th>Diameter (mm)*</th>
<th>Biomass (g)*</th>
<th>% Mycorrhizal roots colonization</th>
</tr>
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<tr>
<td>Without MycoSilvi</td>
<td>-</td>
<td>-</td>
<td>1.83f</td>
<td>1.06i</td>
<td>0.14g</td>
<td>0</td>
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<tr>
<td>+</td>
<td>-</td>
<td>3.75f</td>
<td>1.59h</td>
<td>0.30f</td>
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</tr>
<tr>
<td>-</td>
<td>+</td>
<td>4.07f</td>
<td>1.73g</td>
<td>0.76f</td>
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<td>+</td>
<td>+</td>
<td>1.88f</td>
<td>1.04f</td>
<td>0.21f</td>
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<td>-</td>
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<td>2.70f</td>
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<td>19.37a</td>
<td>1.59h</td>
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<td>-</td>
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<td>2.76a</td>
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<td>MycoSilvi var 3</td>
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<td>12.33dc</td>
<td>2.15de</td>
<td>1.44d</td>
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<td>+</td>
<td>-</td>
<td>11.15d</td>
<td>2.24cde</td>
<td>1.26 de</td>
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<td>+</td>
<td>19.45a</td>
<td>2.63ab</td>
<td>2.14 ab</td>
<td>98</td>
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<td>14.67bc</td>
<td>2.02ef</td>
<td>1.49 ed</td>
<td>78</td>
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</table>

Note: *Values followed by the same letter (s) in the same columns are not significantly different at p<0.05, Duncan’s Multiple Range Test (DMRT)
The positive effects of liming in acid soils for improving plant growth are well documented (Bambara and Ndakidemi 2009; Guo and Huang 2010; Moreira et al. 2010). In this study, soils amended by lime increased plant height, stem diameter and biomass by 123%, 62%, and 444% respectively as compared to those of control plants (Table 1). As reported by Furtini-Neto et al. (2004), liming can increase soil pH and reduces the fixation of P by Al and Fe, stimulates roots growth and finally increase nutrient uptake by plant roots. In our study, liming can increase soil pH, decrease Al and Fe in soil media (Table 2).

The interesting finding from this research is that the lime increased AMF development, as indicated by increasing mycorrhizal roots colonization in all variants of MycoSilvi being applied (Table 1). Consequently, the growth of plants also increased significantly. Interaction between MycoSilvi variant 1 and lime increased significantly plant height, stem diameter and biomass by 847%, 159%, and 1486% respectively as compared to control plants, while MycoSilvi variant 2, when combined with lime, increased significantly plant height, stem diameter, and biomass by 880%, 152%, and 1106% respectively. The same combination with MycoSilvi variants 3 increased significantly plant height, stem diameter and biomass by 965%, 147%, and 1427% respectively. In this study, as shown in Table 2, lime facilitated the increase of soil pH and decrease of Al content in soil growth medium, and such conditions become favorable for root growth which is preferable site for AMF development. Our finding confirms the results reported by Bambara and Ndakidemi (2009), Moreira et al. (2010) and Guo and Huang (2010).

The interaction between MycoSilvi, compost, and lime have been reported by Jayani et al. (2018), who found that plants inoculated with AMF and combined with lime and compost in growth medium, significantly increased height and diameter growth by 148.3 and 155.9% respectively as compared to plants inoculated with MycoSilvi combined with lime alone in soil growth medium. On the contrary, in our results, plants inoculated with MycoSilvi and combined with compost and lime, showed reduced plant height and diameter as compared to plants inoculated with MycoSilvi combined with lime alone in soil growth medium. The extent of decreasing plant height and diameter varied, depending on MycoSilvi being used. When soil growth media were amended with lime and compost, and the plants were inoculated with MycoSilvi variant 1, 2 and 3, the plant heights were reduced by 59.1%, 38.9%, and 32.6% respectively, and diameter growth was reduced by 30.8%, 25.1% and 30.2% respectively, as compared to plants inoculated with MycoSilvi combined with lime alone in soil growth medium. These different results are probably due to different C/N ratio of compost being used. In our research compost has high C/N ratio, namely 42.68, while Jayani et al. (2018) used compost with C/N ratio 25.76. According to Kuzyakov and Xu (2013), organic compost with high C/N ratio may inhibit plant growth due to nutrient competition between decomposer microorganisms and plant roots.

### Relative mycorrhizal dependency and growth response of *Falcatoria moluccana*

The potential role of AMF in restoration of degraded land has been reported by Asmelaash et al. (2016), and the contribution of AMF to plant growth depends on soil fertility. The degree in which a plant is dependent on the mycorrhizal condition to produce its maximum growth or yield, at a given level of soil fertility is called mycorrhizal dependency (Gerdemann 1975). In our research, the degree of mycorrhizal dependency of *F. moluccana* varied depend on the combined treatments (Figure 1). *F. moluccana* inoculated by MycoSilvi variant 1, 2 and 3, without soil amendment had relative mycorrhizal dependency 76%, 90%, and 90% respectively. These results were in agreement with Jha et al. (2012), who found that maximum mycorrhizal dependency of Bamboo species was achieved by plants inoculated with *Acaulospora scrobiculata*.

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**Table 2. Chemical characteristics of soil medium after treatment at the end of experiment**

<table>
<thead>
<tr>
<th>MycoSilvi</th>
<th>Soil ameliorant</th>
<th>Compost</th>
<th>Lime</th>
<th>pH (H₂O)</th>
<th>Al (mg/kg)</th>
<th>Fe (mg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Without MycoSilvi</td>
<td>-</td>
<td>-</td>
<td>4.65</td>
<td>224.54</td>
<td>138.23</td>
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</tr>
<tr>
<td>+</td>
<td>-</td>
<td>4.32</td>
<td>181.97</td>
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<tr>
<td>-</td>
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<td>6.17</td>
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<td>+</td>
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<td>5.93</td>
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<td>41.00</td>
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<td>4.20</td>
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<td>-</td>
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<tr>
<td>+</td>
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<td>+</td>
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<td>4.27</td>
<td>146.42</td>
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<td>3.90</td>
<td>180.37</td>
<td>105.76</td>
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<td>+</td>
<td>-</td>
<td>4.27</td>
<td>146.42</td>
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</table>
followed by *Glomus cerebiforme* and *G. intraradis*. In our results, the maximum relative mycorrhizal dependency of *F. moluccana* was recorded for MycoSilvi variant 3 which contained *Glomus mosseae*, *Acaulospora* sp. and *Gigaspora margarita*, followed by MycoSilvi variant 2 which contained *G. mosseae* and *Acaulospora* sp. (Figure 1).

The degree of relative mycorrhizal dependency was also affected by soil condition as reported by previous researchers (Renuka et al. 2012) who found that the relative mycorrhizal dependency of *Acacia melanoxylon* correlated with the degree of disturbance of the soil which is characterized by unfertile soil. As shown in Figure 1, the relative mycorrhizal dependency of *F. moluccana* inoculated with MycoSilvi variants 1, 2 and 3 and treated with lime were decreased by 66%, 55%, and 64% respectively, indicating that liming effectively increased soil pH and released P to the soil growth medium, and finally plant roots can absorb normally and dependency to mycorrhizae decreased. On the contrary, the addition of compost increased relative mycorrhizal dependency of *F. moluccana* by 88%, 85% and 76% for MycoSilvi variant 1, 2 and 3 respectively, probably due to the C/N ratio of compost used in this study which was quite high.

The responsiveness of plant to mycorrhizal inoculation was called as growth response (Plenchette et al. 1983). Figure 2 presented the growth response of *F. moluccana* as affected by soil amendment. *F. moluccana* was very responsive to MycoSilvi variant 3, followed by MycoSilvi variant 2 and Variant 1.

The difference of mycorrhizal dependency and responsiveness was due to differences in plant species (Tawaraya 2003) and differences in the development of external hyphae and nutrient uptake from the soil (Smith et al. 2000). The addition of lime and compost to soil medium increased soil pH and decreased Aluminium and Fe in the soil (Table 2), and consequently plant roots grow better for absorbing nutrients from the soil and finally decreased the growth responsiveness of *F. moluccana* (Figure 2).

The important findings of this research were that post-mining soil which was characterized by very low soil fertility could be improved for *F. moluccana* plant growth by application of MycoSilvi, lime and compost alone or in combination. To maximize the positive effect of MycoSilvi on plant growth, lime or compost can be added to soil growth medium. Based on our results, MycoSilvi variant containing more than one AMF species was better than those which contain only one AMF species. *F. moluccana* has a high growth response and mycorrhizal dependency on degraded soil. These findings imply the prospective use of MycoSilvi and soil ameliorant for improving plant growth in unfertile soil medium, including soil in post-mining area, and support the success of rehabilitation of degraded land area.

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**REFERENCES**


Widyati E. 2006. Bioreminder of Former Coal Mining Land with Paper Industry Sludge to Spur Land Revegetation [Dissertations] IPB University, Bogor. [Indonesian]